

OPEN ACCESS POLICY

Indian Journal of Veterinary and Animal Sciences Research is an open access journal. All articles published by the Indian Journal of Veterinary and Animal Sciences Research (IJVASR) are freely and permanently accessible online at <http://www.tanuvas.tn.nic.in/ijvasr/ijvasr-articles.html>

INDIAN JOURNAL OF VETERINARY AND ANIMAL SCIENCE RESEARCH
(Bi-monthly)

EDITORIAL BOARD

Chief Editor

Dr.S.Thilagar

Vice- Chancellor

Tamil Nadu Veterinary and Animal Science University
Madhavaram Milk Colony, Chennai – 600 051

Editor

Dr.T.J.Harikrishnan

Director of Research
Tamil Nadu Veterinary and Animal
Science University
Madhavaram Milk Colony,
Chennai – 600 051

Associate Editor

Dr.S.Balasubramanian

Director of Clinics,
Tamil Nadu Veterinary and Animal
Science University
Madras Veterinary College Campus,
Vepery, Chennai- 600 007

Members

Dr.Geetha Ramesh

Professor and Head
Dept.of Veterinary Anatomy & Histology
Madras Veterinary College,
Chennai – 7

Dr.C.Ramani

Professor
Dept.of Vet. Surgery and Radiology
Madras Veterinary College,
Chennai – 7

Dr.H.Gopi

Professor and Head
Post Graduate Research Institute in
Animal Science
Kattupakkam

Dr.Y.Krishnamohan Reddy

Professor and Head
Vaccine Research Centre-Viral Vaccines,
Centre for Animal Health Studies,
Madhavaram Milk Colony,
Chennai – 600 051

Dr.S.C.Edwin

Professor and Head
Dept.of Livestock Production and
Management, VC&RI,
Tirunelveli

Dr.B.Dhanalakshmi

Professor and Head,
Dept.of Livestock Products Technology
(Dairy Science),
Madras Veterinary College, Chennai – 7

Dr.V.Leela

Professor and Head,
Dept.of Veterinary Physiology
Madras Veterinary College,
Chennai – 7

Dr.R.Sridhar

Professor and Head
Dept.of Veterinary Pathology
Veterinary College and Research
Institute, Tirunelveli

Dr.O.R.Sathyamoorthy

Professor, Office of the Registrar,
Tamil Nadu Veterinary and Animal
Science University,
Madhavaram Milk Colony,
Chennai – 600 051.

Dr.R.K.Kanimozhi

Assistant Professor
Office of the Directorate of Distance
Education (DDE)
Nandanam, Chennai- 600 035.

INDIAN JOURNAL OF VETERINARY AND ANIMAL SCIENCES RESEARCH
(Bi-monthly)

INTERNATIONAL EDITORIAL ADVISORY BOARD

Dr. Yung-Fu Chang

Director, Infectious Disease Research Laboratory
Animal Health Diagnostic Center
Professor
Department of Population Medicine and
Diagnostic Sciences
C1-114, Vet Medical Center
College of Veterinary Medicine
Cornell University, Ithaca
New York 14853-5786, USA

Dr. John Gilleard, BVSc, Ph.D, Dip EVPC, MRCVS

Director of Research
Dept. of Comparative Biology and
Experimental Medicine
Faculty of Veterinary Medicine
University of Calgary
3330, Hospital Drive NW
Calgary
Alberta
Canada

Dr. Puliur S. Mohankumar, B.V.Sc., Ph.D.

Professor
Department of Biomedical Sciences & Diagnostic Imaging
College of Veterinary Medicine
University of Georgia
Athens, GA 30602, USA

Dr. Damer Blake, MSc, Ph.D, PGC Vet Ed, FHEA

Lecturer in Molecular Parasitology
Dept. of Pathology and Pathogen Biology
The Royal Veterinary College
University of London
Hatfield, Herts AL 9 7TA
United Kingdom

Prof. Dr. Terry Spithill

Co-Director of AgriBio
The Centre for AgriBio Science
Faculty of Science, Technology & Engineering
School of Life Sciences,
La Trobe University
5, Ring Road, Bundoora
Melbourne Victoria 3086
Australia

Attention Contributors

The Editorial Board of Indian Journal of Veterinary and Animal Sciences Research has decided to collect Rs.500/- (Rupees Five hundred only) as processing fee in accordance with the order of Registrar, TANUVAS-(U.S.O.No.500601/G4/2016 Proc. No. 5639/G4/2016 dt 3.5.2016),from the authors at the time of submission of articles for publication in the Journal. This would help the authors to hasten the publication of their articles without any delay.

Hence, the corresponding author is requested to draw a demand draft for Rs.500/- in favour of “The Editor, IJVASR & Director of Research, TANUVAS,Chennai-600051”along with the manuscript during submission. The articles may be addressed to the Associate Editor, IJVASR & Director of Clinics, Madras Veterinary College Campus, Chennai-7. The authors are also requested to mention their contact phone number and E-mail address.

Chief Editor

Review articles invited from eminent Scientists

The Editorial Board of Indian Journal of Veterinary and Animal Sciences Research invites review articles from eminent research scientists in the field of Veterinary and Fisheries Sciences, on the latest/ current topics of interest for publication in the Journal. The review article (both hard and soft copy) may please be sent to the Editor/Associate Editor, Indian Journal of Veterinary and Animal Sciences Research for publication.

Chief Editor

INDIAN JOURNAL OF VETERINARY AND ANIMAL SCIENCES RESEARCH
(Formerly Tamil Nadu Journal of Veterinary and Animal Sciences)

This Journal is published bi-monthly by Tamil Nadu Veterinary and Animal Sciences University, Chennai in February, April, June, August, October and December of every year.

1. Annual Subscription (Inland) - Rs.500/- (Rupees Five hundred only)
2. Life Membership (Inland) - Rs.3000/- (Rupees Three thousand only)
(for 10 years)
3. Processing fee (Inland) - Rs.500/- (Rupees five hundred only)
4. Annual Subscription (Foreign) - US \$50/- (Fifty US Dollars only)
5. Life Membership (Foreign) - US \$250/- (Two hundred and fifty US Dollars only)
6. Processing fee (Foreign) - US \$10/- (Ten US Dollar only)

Subscriptions are payable in advance and the subscription rates are inclusive of postal charges.

Demand draft is to be drawn in favour of “The Chief Editor, IJVASR & Director of Research, TANUVAS, Chennai - 51. Advance payment of annual subscription is solicited for uninterrupted supply of the journal.

The first / corresponding authors are requested to inform their email addresses and contact numbers while submitting manuscripts to this journal.

Chief Editor

INDIAN JOURNAL OF VETERINARY AND ANIMAL SCIENCES RESEARCH
(Formerly Tamil Nadu Journal of Veterinary and Animal Sciences)

Vol. 46

November - December 2017

No. 6

Review article

1. CONTRAST ENHANCED ULTRASOUND IN SMALL ANIMAL PRACTICE
Dr. Sethupandian Prathaban

Full length articles

2. EVIDENCE OF SHARED GENOMIC SEGMENT 6 BETWEEN BLUETONGUE VIRUS SEROTYPES 1 AND 2
Y. Vishuvaradhan Reddy, B. Susmitha, Y. Narsimha Reddy, D. Sreenivasulu, Kalyani Putty, K.S.R. Siva Sai, Pavuluri Panduranga Rao, Nagendra R Hegde
3. DEVELOPMENT OF GHEE RESIDUE CANDY INCORPORATED WITH ORANGE PEEL
Ananthakumar.K.V and B.Dhanalakshmi
4. INFLUENCE OF HOUSING SYSTEM ON CUMULATIVE FEED CONSUMPTION AND FEED CONVERSION RATIO OF BROILERS FED WITH DIFFERENT LEVELS OF ENERGY AND PROTEIN
K. Rajendran, K. Mani, P. Shamsudeen, V. Ramesh Saravana Kumar and A. Natarajan

Short Communication

5. ANTIMICROBIAL EFFECT OF PSIDIUM GUAJAVA, MANGIFERA INDICA AND PUNICA GRANATUM EXTRACTS AGAINST BOVINE MASTITIS PATHOGEN
Silambarasan D, Amsaveni S, Kalaiselvi L and Ramesh S
6. EVALUATION OF POSTERIOR CAPSULAR OPACIFICATION IN DOGS – A REVIEW OF 12 CASES
Gokilavani. M, C. Ramani, Mohamed Shafuzama, S. Ramesh
7. EFFECT OF SILK FIBROIN LOADED SILVER NANOPARTICLES GEL ON WOUND HEALING IN THREE LABRADOR DOGS
T.S. Premavathy, Mohamed Shafuzama, R.Siva Shankar, Mala Shammi, S. Hemalatha
8. GRANULOSA CELL TUMOUR ALONG WITH PYOMETRA IN A GERMAN SHEPHERD DOG
B. Bibin Becha, Sunita Behera, M. Navya and G. Sudha

CONTRAST ENHANCED ULTRASOUND IN SMALL ANIMAL PRACTICE

Dr. Sethupandian Prathaban

Professor and Head, Department of Veterinary clinical Medicine (Retd),
Madras Veterinary College, Chennai - 600 007

Conventional grey scale ultrasonography are non-invasive techniques that, by virtue of their cross sectional nature and ability to detect blood flow, have had a considerable impact on the imaging assessment of small animal patients. Despite these advantages, the broadly similar attenuation properties of normal and diseased tissues may, in certain instances, limit identification and characterisation of pathology. Furthermore, the weak echoes and physical characteristics of blood flow restrict the various Doppler techniques to the assessment of macrovascular structures. Ultrasound contrast agents (UCAs) overcome this limitation by substantially increasing the signal generated within the tissues in which they accumulate and contrast. Furthermore by acting as blood pool agents, UCAs demonstrate microvasculature –thus permitting perfusion studies and functional imaging (King 2006).

Contrast ultrasound: Contrast agents have been integral in all imaging modalities except US. The reasons for this limitation in ultrasound imaging is because of the cost of media and need for intravenous pool access. Unlike MR and CT contrast media, ultrasound contrast agents, are pool agents. These contrast agents are microscopic bubbles made from an outer shell and central

core of gas. There are limited numbers of manufacturers of ultrasound contrast media. Older first generation contrast agents use air in the core (Lenovist). Air is not a potent medium for emission of ultrasound signal and results in poor images. Second generation contrast media utilize inert gases in the core, which provide more non-linear effects. The first second generation contrast media (Optison) used human albumin in the shell. This had obvious limitations in veterinary medicine for immunological concerns.

Second generation contrast agents utilize an immunologically inert lipoprotein shell and more efficient inert gas in the lumen. The range of bubble sizes are predominantly small (most less than 10 micrometers), pass through the pulmonary circulation unaffected and can be imaged in the small vessels of all organs currently being imaged in the diagnostic veterinary imaging. Contrast media currently recommended for the veterinary clinical applications are Definity (USA) and Sonovue (Canada). Both are very safe in dogs and no side effects have been reported. The bubbles of Definity are more concentrated and rigorous than Sonovue providing an advantage for clinical characterisation or detection of lesions, such as liver nodules. For experimental studies Sonovue is advantageous.

New contrast agents are available like Targestar is a blood pool agent that demonstrates great promise because of high resonant frequency and improved stability of the reconstituted bubbles. Reconstituted Targestar is stable with refrigeration for weeks and months. This agent has the ability conjugate to an active coupling molecule. This is a very exciting new area of research called molecular imaging. Conjugation of ligands to a blood pool US contrast agent may provide an opportunity to couple monoclonal antibodies, various mediator proteins or possible therapeutics (Correas *et al.*, 2009).

Practical advantage of using contrast enhanced ultrasound includes the relatively low cost of contrast ultrasound examinations: comparable sensitivity to computed tomography and magnetic resonance imaging for detecting the metastatic disease and the absence of ionising radiation.

Limitation is that the keeping quality is very short and should be used within hours after its reconstruction.

CLINICAL APPLICATION

Liver: UCA demonstrates affinity for certain organs, such as liver in which they persist. The reason for the accumulation is not very clear, but is suspected to be associated with accumulation within the hepatic sinusoids or incorporation within Kupffer cells. As a result, three phases of liver enhancement –arterial, early and delayed portovenous phases– can be recognised. The clinical utility of the UCA is particularly related to the perfusion in

the later two phases. A continuous low MI imaging technique is usually employed in contrast enhanced examinations of the liver. The use of contrast ultrasonography in dog and cat livers has been widely reported in the localisation and characterisation of nodular hepatic lesions–in particular the differentiation of benign primary nodular disease from metastatic or primary hepatic malignancy. In general benign lesions including lipid granulomas, tend to demonstrate variable enhancement during the arterial and early portovenous phases (hypo, iso or hyperechoic) with similar enhancements to liver during normal liver during the late portal phase.

Malignant lesions characteristically show a rapid and complete contrast washout and appear hypoechoic, compared to the enhanced liver, during the late portovenous phase. These characteristics are related to tumoral neo angiogenesis from the hepatic artery and the absence of normal hepatic sinusoidal architecture. Hemangiosarcoma metastases, particularly relevant in veterinary patients, may not demonstrate enhancement at any stage. The imaging characteristics during the early phases of liver enhancement generally appear to be of little clinical significance in the assessment of metastatic phase. Although macrovascular patterns, such as the basket pattern associated with hepatocellular carcinoma and the spoke wheel pattern typical of nodular hyperplasia, may be of significance in humans with primary hepatic masses, the validity of extrapolating these features to the dog and cat is uncertain. Diagnosis of large complex or lobar masses should be done with caution, as these may have areas of haemorrhage or

necrosis that confound the diagnosis. Careful correlation with fundamental images and anticipated histopathological findings is necessary, as predominantly cystic lesions may demonstrate rapid wash out and non enforcement of haematomas may mimic malignant disease.

Contrast ultrasonography of the liver should be employed primarily to assess lesion margination and extent and to improve lesion detection in veterinary clinical practice. Isoechoic or poorly marginated solid components of masses with larger cystic or necrotic regions may not be recognised on fundamental imaging. Contrast enhanced ultrasonography may then be used to guide biopsy to avoid sampling non-representative regions of such mass lesions. Equally iso echoic or poorly marginated metastatic nodules may be difficult to recognise on fundamental imaging and contrast enhanced ultrasound is reported to increase the number of metastases identified. These factors particularly relevant where exploratory surgery is being considered. The findings of the contrast enhanced examination and aspirate or biopsy results should not be interpreted in isolation, but considered together to determine if they are representative. Two or three UCA boluses are usually required to ensure a satisfactory assessment of all liver segments. The quality of the later examinations is usually superior due to UCA persistence in the liver. The technique is dependant on operator experience and all studies should be acquired as cine loops for review.

Salweiet *al.* (2005) described the use of ultrasound in the diagnosis of portosystemic

shunts (PSS). Shunting vessels are not recognised as such, but the generation of time –intensity curves of hepatic perfusion demonstrate a shorter time to peak liver enhancement when compared to normal dogs, due to the marked increase in hepatic arterialisation associated with PSS.

Spleen: Spleen demonstrates an early arterial phase of enhancement with a patchy or mottled appearance. The late paranchymal phase results in more homogenous enhancement and should be used to asses any changes. Contrast ultrasonography has been reported to be useful to discriminate between benign and malignant lesions. Benign lesions are characterised by enhancement at a rate and intensity similar to that of a normal spleen. Malignant tumors are characterised by a hypoechoic appearance during the late paranchymal phase, with rapid enhancement (wash in) a feature of lymphoma and soft tissue sarcomas. Extensive non-perfused regions during all phases are a feature of haemangiosarcoma. The diagnosis of lymphoma is usually facilitated by identifying concurrent lymphadenopathy or organomegally and cytology is usually diagnostic. Therefore contrast enhanced ultrasound should reserved for those cases with disease limited to spleen (suspected to stage four disease) and or/ to support equivocal cytological findings. Differentiation between haematoma and haemangiosarcoma is not possible (Ivan *et al.*, 2009). UCAs may be useful to confirm and differentiate splenic torsion or infarction, from splenitis infiltrative disease, where the conventional B mode findings are not typical for the appearance of vascular compromise.

Pancreas: The use of contrast enhanced ultrasound has been reported in the characterisation of pancreatic disease in humans. The utility and relevance of the technique in canine pancreatitis has not been demonstrated, but may be of value in identifying extensive or irreversible pancreatic necrosis. Contrast ultrasound of the diseased pancreas in the demonstrated increased perfusion and vascularity compared to normal cats according to one study (Rademacher *et al.*, 2008).

Differentiation between inflammatory benign nodular changes and neoplastic disease is possible using this technique. Insulinoma in common with other neuroendocrine tumors, may demonstrate strong enhancement during the arterial phase, with rapid washout of contrast media. Contrast enhanced ultrasonography of the superficial lymph nodes has been reported to better define the distribution of vessels, which may characterise lesion morphology (Salwei *et al.* 2005). This may assist in staging of disease or demonstrate sentinel lymph node involvement.

CEUS has been used extensively in fields such as cardiology and hepatology. This technique is promising for studies for kidney structure and function because of its safety profile and its ability to provide information on single kidney as well as regional blood flow. It has been used to determine changes in renal function in response to physiologic and pharmacologic interventions and to study disease such as renal artery stenosis and post kidney transplantation (Kalantarinia and Okusa 2007).

Contrast agents can increase echogenicity within and around the altered parenchyma, thus optimizing Doppler evaluation and consequently the diagnosis of prostate gland disease in dogs (Bigliardi and Ferrari, 2010).

Contrast enhanced ultrasound is a valuable, minimally invasive technique particularly suited to the identification and characterisation of liver and splenic nodular lesions, organs and diseases. This technique may assist in identifying suitable areas for tissue sampling. The technique's value for the evaluation of other organs and diseases and more work is progressing in this area. The requirements of specialised equipment, transducers and software, the dependence of operator experience and the cost of the contrast ultrasound media must be recognised as significant limitations.

REFERENCES

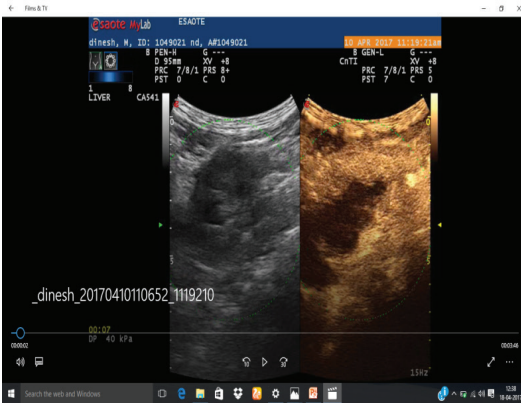
- Bigliardi and Ferrari (2011) Contrast enhanced ultrasound of the normal canine prostate gland, *Veterinary Radiology and Ultrasound*, 52:107-110.
- Correas, J.M., Claudon, M., Tranquart, F. and Helenon, A.O. (2006) The Kidney: Imaging with microbubble contrast agents, *Ultrasound Q* 22:53-66
- Ivan, M., Long, F. and Seiler, G.S. (2009) Contrast harmonic ultrasonography of splenic masses and associated liver nodules in dogs, *J. Amer. Vet. Med. Ass* 234:88-94

Kalantarinia,K., and Okusa,M.D (2007)
Ultrasound contrast agents in the
kidney Function in Health and Disease
, Drug Discov Today Dis Mech 4:153-
158

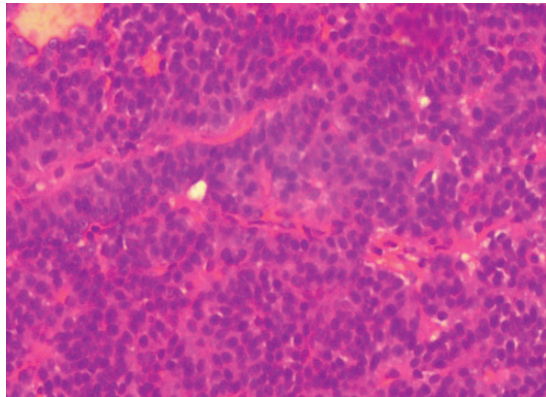
King A.M., (2006) Development advances
and application of diagnostic
ultrasound in animals, The Veterinary
Journal 171:408-420

Rademacher N,Ohlerth S, Scharf G,
Laluhova D, and Ross M (2008)
Contrast enhanced power and colour
Doppler ultrasonography of the
pancreas in healthy and diseased cats,
J.Vet.Int. Med 22{1310-1316.

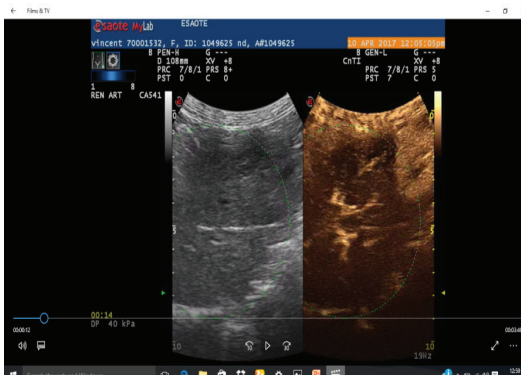
Salwei RM,O'Brien RT, and Matheson
JS(2005) Cacterisation of
lymphomatous lymph nodes in dogs
using contrast harmonic and power
Doppler ultrasound ,. Vet.Radiol.
Ultrasound 46:411-416.



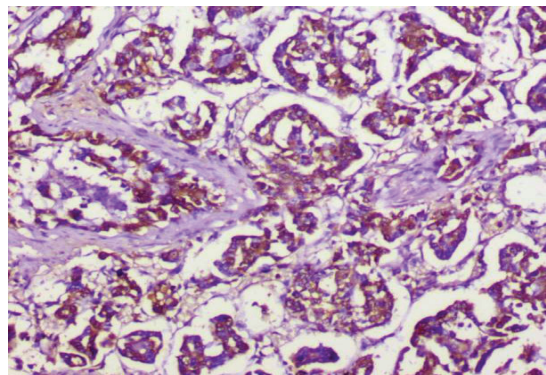
**Peripheral enhancement Pattern –
Ductal tubular adenocarcinoma of
Liver**



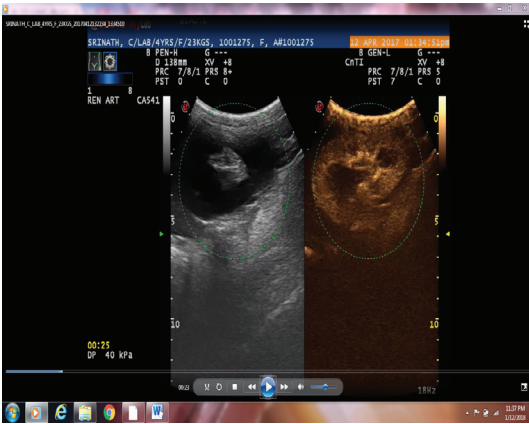
**Liver-HP-Metastatic Ductal
adeno carcinoma. H&E 200 x**



**Radial enhancement –
Cholangiocellular carcinoma**



**Liver-IHC- Cytokeratin -Cholngio cellular
carcinoma. Diffuse intense cytoplasmic signal.
DAB 200x**



CEUS of Kidneys: Filling of Renal cortex with contrast agent

CEUS of Kidneys: Filling of Renal cortex with contrast agent

Method of preparation of sonovue

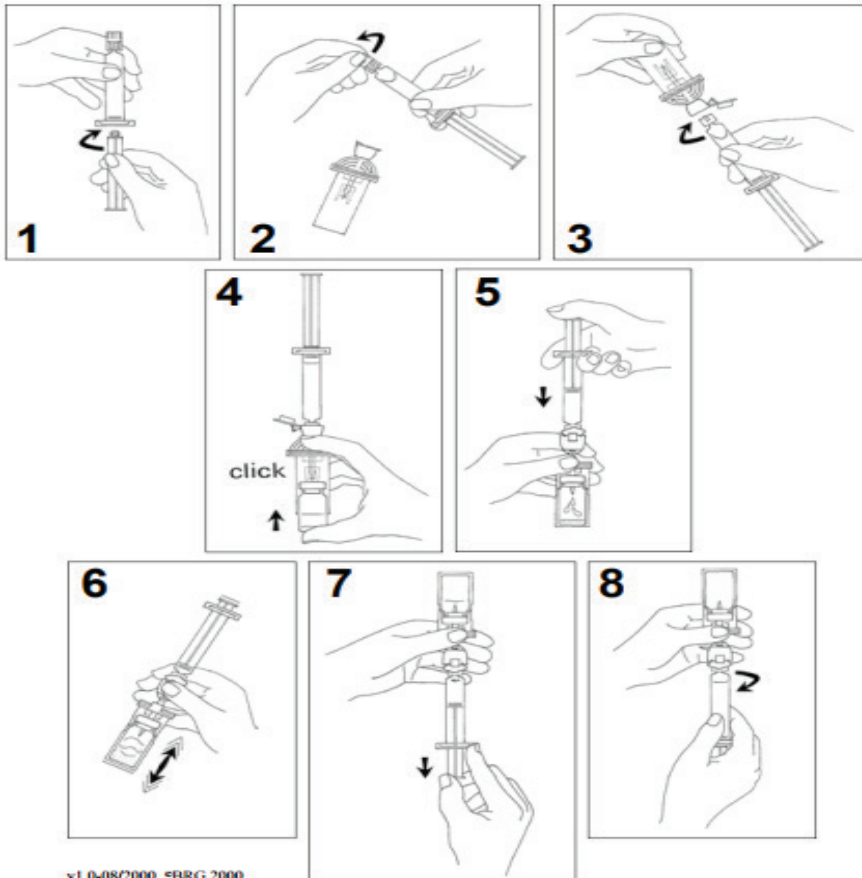


Image from product catalogue

1. Connect the plunger rod by screwing it clockwise into the syringe.
2. Open the MiniSpike transfer system blister and remove syringe tip cap.
3. Open the transfer system cap and connect the syringe to the transfer system by screwing it in clockwise.
4. Remove the protective disk from the vial. Slide the vial into the transparent sleeve of the transfer system and press firmly to lock the vial in place.
5. Empty the contents of the syringe into the vial by pushing on the plunger rod.
6. Shake vigorously for 20 seconds to mix all the contents in the vial to obtain a white milky homogeneous liquid.
7. Invert the system and carefully withdraw SonoVue into the syringe.
8. Unscrew the syringe from the transfer system.
 - Do not use if the liquid obtained is clear and/or if solid parts of the lyophilisate are seen in the suspension.
 - SonoVue should be administered immediately by injection into a peripheral vein for use in echocardiography and in vascular Doppler imaging
 - If SonoVue is not used immediately after reconstitution the microbubble dispersion should be shaken again before being drawn up into a syringe.
 - Chemical and physical stability of the microbubble dispersion has been demonstrated for 6 hours.

EVIDENCE OF SHARED GENOMIC SEGMENT 6 BETWEEN BLUETONGUE VIRUS SEROTYPES 1 AND 2

Y. Vishuwardhan Reddy¹, B. Susmitha¹, Y. Narsimha Reddy², D. Sreenivasulu³,
Kalyani Putty², K. S. R. Siva Sai⁴, Pavuluri Panduranga Rao^{1,*},
Nagendra R Hegde^{1,5}

ABSTRACT

Bluetongue virus (BTV) belongs to the genus *Orbivirus* of family *Reoviridae*. Twenty seven serotypes of BTV have so far been recognized. The genome of BTV consists of ten double-stranded segmented RNA which can reassort independently during mixed infections. The outer capsid proteins VP2 and VP5, coded by segments 2 and 6, are the major determinants of serotype, and serotype-dependent linkage between these segments has been observed. Natural and *in vitro* reassortants of segment 2 and 6 of different serotypes of BTV have been reported earlier. Here we report a BTV isolate which is a natural reassortant of segment 2 and 5 of BTV-1 and -2.

Key words: Bluetongue virus, Serotype, Australasian topotype, Segment reassortment, Shared segment 6

INTRODUCTION

Bluetongue (BT) is endemic in most of the tropical and some of the subtropical areas. The disease is caused by bluetongue virus (BTV) belonging to genus *Orbivirus*, family *Reoviridae*. The BTV structure is a triple layered icosahedron. The innermost subcore is a cage formed by the viral protein (VP) VP3. The subcore encases the viral RNA which is associated with VP1, VP4 and VP6, together forming the transcriptase

complex (Mertens & Diprose, 2004). The middle, outercore layer is composed of VP7. Finally, the outermost, outer capsid is formed by VP2 and VP5 proteins (Roy, 2005). The genome of the virus consists of ten segments of double-stranded RNA designated as Seg-1 to -10 in the order of decreasing molecular size, and coding respectively for VP 1 (Seg-1), VP2 (Seg-2), VP3 (Seg-3), VP4 (seg-4), NS1 (Seg-5), VP5 (Seg-6), VP7 (Seg-7), NS2 (Seg-8), VP6 , NS4 (seg-9) and NS3, NS2a, NS5

¹Ella Foundation, Hyderabad-500078

²College of Veterinary Science, SPVNR Veterinary University, Hyderabad-500030

³College of Veterinary Science, SV Veterinary University, Tirupati-517502

⁴Sreenidhi Institute of Science and Technology, Hyderabad-501301

⁵Present address: National Institute of Animal Biotechnology, Hyderabad-500049

*Corresponding Author: P. Panduranga Rao, Ella Foundation Genome Valley, Turkapally, Shameerpet Mandal, Hyderabad-500078, E-mail: raopp@ellafoundation.org

(Seg-10) proteins (Sung & Roy, 2014). The genomic RNA segments can reassort independently and can generate several variants of BTV during mixed infections. Genome segment reassortment is thought to be one of the important mechanisms of BTV evolution and fitness (Niedbalski, 2013; Nomikou et al., 2015; Shaw et al., 2013a).

BTV exists in 27 serotypes. Although, VP2 and VP5 are serotype determining proteins (Mertens et al., 1989), it has been shown by us and others that VP5 can be shared among closely related serotypes (Rao et al., 2017; Shaw et al., 2013a). Natural reassortants of segment 2 and 6, coding for VP2 and VP5, respectively, has been observed between BTV-1 and -8. Furthermore, *in vitro* reassortment of segment 2 and 6 between BTV-20 and -21 has also been reported earlier. In this study, we describe the investigation of a BT outbreak and characterization of the associated BTV-1 and -2 serotype isolates. We found and report possible reassortment and sharing of segment 6 between those serotypes.

MATERIALS AND METHODS

Isolation of the viruses

BTV from field samples was isolated by passaging in embryonated chicken eggs (ECE) followed by passaging in BHK-21 (Clavijo et al., 2000). Briefly, heparinized blood samples were collected aseptically from sheep showing BT-like symptoms, and used to inoculate ECE. Triturates of ECE showing hemorrhages were used to infect BHK-21 cells, and observed for cytopathic

effect (CPE) characteristic of BTV infection. Details of viruses used during this study are given in Table 1. Culture supernatants from cells showing CPE were titrated on BHK-21, and the Reed and Muench method (Reed & Muench, 1938) was applied to calculate the 50% tissue-culture infectious dose (TCID₅₀).

RNA isolation and sequencing

BHK-21 cells were infected with different BTV isolates at a multiplicity of infection (MOI, i.e., TCID₅₀ / cell) of 0.01 and incubated at 37°C until CPE was observed. Total RNA was isolated from cell lysates using the MPGIT solution (Gauthami et al., 2015) and cDNA was synthesized. Sequencing library was prepared and cDNA was sequenced by “sequencing by synthesis” (SBS) on Illumina HiSeq (Rao et al., 2013). Briefly, RNA was fragmented into small pieces, reverse transcribed, multiplexing adapters were ligated to the cDNA ends and PCR was performed using primers complimentary to the adapters. PCR products were purified, diluted to obtain 100-1000x coverage of the genome, heat denatured and hybridized to oligonucleotides immobilized on flow cell. Paired end sequencing was carried out, adapter sequences were deleted and reads were assembled using Velvet 1.1.01 (Zerbino & Birney, 2008) or Bowtie1.0 (Langmead et al., 2009). BTV-specific reads were identified by aligning them to reference sequences and the sequences were submitted to GenBank (Table 1).

Sequence analysis

Segment 2 sequences were analysed to identify the serotype of the virus. For

phylogenetic analysis, coding sequences of segment 6 of global isolates of BTV-1 and BTV-2 were aligned, and Maximum Likelihood (ML) tree was constructed with TN93+G+I as nucleotide substitution model with 1000 Bootstrap replications using MEGA 5.03 suite (Tamura et al., 2011).

RESULTS AND DISCUSSION

Three isolates of BTV-1 (BTV01IND2007, BTV01IND2010/01, BTV01IND2010/02) and two isolates of BTV-2 (BTV02IND1993, BTV02IND2010) were sequenced using SBS method and Seg-2 and Seg-6 sequences coding for VP2 and VP5 were analysed. Sequence homology of Seg-2 indicated that all the BTV-1 isolates belonged to the Australasian topotype with sequence similarity of more than 87%. Similarly, Seg-2 of BTV-2 isolates also belonged to Australasian topotype with sequence similarity of more than 89%. Seg-6 sequences of all the Indian isolates of BTV-1 were similar (more than 90%) to that of other Australasian BTV-1 sequences. While Seg-6 of BTV02IND2010 had high sequence similarity (>99%) with one of the Indian isolates of BTV-2 isolated in 1982 and Australasian viruses (>91%), Seg-6 of BTV02IND1993 had very high sequence similarity with Indian isolates of BTV-1 (>99%) and Australasian viruses (>90%).

Phylogenetic analysis of Seg-6 of BTV-1 and -2 from different parts of the world indicate that this segment did not cluster monophyletically according to serotype. BTV-1 and 2 of African and American viruses formed monophyletic groups and are more related to each other than their relation with Seg-6 of BTV-1 and

-2 serotype viruses of Australasian origin. The available data indicate that Seg-6 of Australasian BTV-1 and -2 also clustered as per their serotype, except in the case of BTV02IND1993.

BTV is a segmented RNA virus and all the genomic segments of BTV can reassort independently during mixed infections, leading to the generation of huge number of variants (Niedbalski, 2013; Nomikou et al., 2015; Shaw et al., 2013a). Widespread genomic reassortment followed by selection may be one of the important evolutionary mechanisms of BTV. Although all the segments can reassort independently, there seems to be some level of linkage between Seg-2 and -6 coding for VP2 and VP5 the outer coat proteins (Rao et al., 2017; Shaw et al., 2013a). cursory analysis of hundreds of BTV isolates from different parts of the world indicates that VP5 is also a serotype determining protein, and with some exceptions, there is a linkage between Seg-2 and -6 and serotypes. Isolation of a natural reassortant Seg-2 and -6 of BTV-8 and -1 has been reported and Seg-2 and -6 reassortants of these serotypes could be developed using reverse genetic approaches (Shaw et al., 2013a). Similarly, generation of reassortants between BTV-20 and -21 has been reported (Cowley & Gorman, 1989). Here, we report a natural reassortant of BTV-1 and -2 with Seg-2 of BTV-2 and Seg-6 of BTV-1 isolated in 1993. On the other hand, the BTV-2 isolates of 1982 and 2010 are different from this isolate and contain natural BTV-2 specific Seg-2 and -6. Isolation of a natural reassortant of Seg-2 and -6 of BTV-2 and -1 suggests that Seg-6/VP5 can be shared between serotypes 1 and 2, and supports the

hypothesis that VP5 can be shared among closely related serotypes.

Phylogenetic analysis of Seg-6 of BTV-1 and -2 indicate that Seg-6 of western topotypes (African, American) cluster monophyletically, whereas Seg-6 of eastern BTV-1 and -2 probably evolved independently. The isolation of natural reassortant of Seg-2 and -6 of these serotypes also supports the possibility of independent evolution of Seg-6 of BTV-1 and -2 in Australasia. Earlier work indicates that the reassortants are immunologically different from their parent strains (Cowley & Gorman, 1989; Shaw et al., 2013a), and may play a role in escaping host immunity to some extent, in a manner similar to antigenic shift observed with influenza viruses.

To conclude, a novel BTV with reassorted serotype determining segments (Seg-2 and -6) from two different serotypes of BTV i.e., BTV-1 and -2, was identified. Immunological characterization of these viruses may illuminate the role of antigenic shift in epidemiology of BT.

ACKNOWLEDGEMENT

We are thankful to all the veterinarians who alerted us of outbreaks and assisted us in the collection of clinical samples. Funding: This work was partially supported by the Department of Biotechnology, Government of India [grant numbers BT/IN/Indo-UK/FADH/46/SM/2013].

REFERENCES

- Boyle, D.B., Bulach, D.M., Amos-Ritchie, R., Adams, M.M., Walker, P.J., Weir, R. (2012). Genomic sequences of Australian bluetongue virus prototype serotypes reveal global relationships and possible routes of entry into Australia. *Journal of Virology* 86: 6724-6731
- Caporale, M., Wash, R., Pini, A., Savini, G., Franchi, P., Golder, M., Patterson-Kane, J., Mertens, P., Di Gialleonardo, L., Armillotta, G. et al., (2011). Determinants of bluetongue virus virulence in murine models of disease. *Journal of Virology* 85: 11479-11489
- Clavijo, A., Heckert, R.A., Dulac, G.C., Afshar, A. (2000). Isolation and identification of bluetongue virus. *Journal of Virological Methods* 87: 13-23
- Cowley, J.A., Gorman, B.M. (1989). Cross-neutralization of genetic reassortants of bluetongue virus serotypes 20 and 21. *Veterinary Microbiology* 19: 37-51
- Gauthami, S., Meena, K., Kumar, D., Uma, M., Reddy, Y., Rao, P.P., Hegde, N.R. (2015). MPGIT: A concentrated monophasic phenol-guanidinium isothiocyanate reagent for isolation of viral RNA. *Indian Journal of Biotechnology* 14: 376-381
- Gould, A.R., Pritchard, L.I. (1988). The complete nucleotide sequence of the outer coat protein, VP5, of the Australian bluetongue virus (BTV) serotype 1 reveals conserved and non-

- conserved sequences. *Virus Research* 9: 285-292
- Langmead, B., Trapnell, C., Pop, M., Salzberg, S.L. (2009). Ultrafast and memory-efficient alignment of short DNA sequences to the human genome. *Genome Biology* 10: R25
- Lorusso, A., Marcacci, M., Ancora, M., Mangone, I., Leone, A., Marini, V., Cammà, C., Savini, G. (2014). Complete genome sequence of bluetongue virus serotype 1 circulating in Italy, obtained through a fast next-generation sequencing protocol. *Genome announcements* 2: e00093-00014
- Maan, S., Maan, N.S., Ross-smith, N., Batten, C.A., Shaw, A.E., Anthony, S.J., Samuel, A.R., Darpel, K.E., Veronesi, E., Oura, C.A. et al., (2008). Sequence analysis of bluetongue virus serotype 8 from the Netherlands 2006 and comparison to other European strains. *Virology* 377: 308-318
- Maclachlan, N.J., Wilson, W.C., Crossley, B.M., Mayo, C.E., Jasperson, D.C., Breitmeyer, R.E., Whiteford, A.M. (2013). Novel serotype of bluetongue virus, western North America. *Emerging Infectious Diseases* 19: 665-666
- Mecham, J.O., Johnson, D.J. (2005). Persistence of bluetongue virus serotype 2 (BTV-2) in the southeast United States. *Virus Research* 113: 116-122
- Mertens, P., Pedley, S., Cowley, J., Burroughs, J., Corteyn, A., Jeggo, M., Jennings, D., Gorman, B. (1989). Analysis of the roles of bluetongue virus outer capsid proteins VP2 and VP5 in determination of virus serotype. *Virology* 170: 561-565
- Mertens, P.P., Diprose, J. (2004). The bluetongue virus core: a nano-scale transcription machine. *Virus Research* 101: 29-43
- Niedbalski, W. (2013). The evolution of bluetongue virus: genetic and phenotypic diversity of field strains. *Pol J Vet Sci* 16: 611-616
- Nomikou, K., Hughes, J., Wash, R., Kellam, P., Breard, E., Zientara, S., Palmarini, M., Biek, R., Mertens, P. (2015). Widespread Reassortment Shapes the Evolution and Epidemiology of Bluetongue Virus following European Invasion. *Plos Pathology* 11: e1005056
- Rao, P.P., Hegde, N.R., Singh, K.P., Putty, K., Hemadri, D., Maan, N.S., Reddy, Y.N., Maan, S., Mertens, P.P.C. (2017). Bluetongue: Aetiology, Epidemiology, Pathogenesis, Diagnosis and Control. In *Emerging and Re-emerging Infectious Diseases of Livestock*, Bayry J. (ed), pp 3-54. Cham: Springer International Publishing
- Rao, P.P., Reddy, Y.N., Ganesh, K., Nair, S.G., Niranjana, V., Hegde, N.R. (2013). Deep sequencing as a method of typing bluetongue virus isolates. *Journal of Virological Methods* 193: 314-319
- Reddy, Y.V., Krishnajyothi, Y., Susmitha, B., Devi, B.V., Brundavanam, Y., Gollapalli, S.R., Karunasri, N.,

- Sonali, B., Kavitha, K., Patil, S.R. et al., (2016). Molecular Typing of Bluetongue Viruses Isolated Over a Decade in South India. *Transboundary and Emerging Diseases* 63: e412-418
- Reed, L.J., Muench, H. (1938). A simple method of estimating fifty per cent endpoints. *American Journal of Epidemiology* 27: 493-497
- Roy, P. (2005). Bluetongue virus proteins and particles and their role in virus entry, assembly, and release. *Virus Structure And Assembly* 64: 69-114
- Shaw, A.E., Ratinier, M., Nunes, S.F., Nomikou, K., Caporale, M., Golder, M., Allan, K., Hamers, C., Hudelet, P., Zientara, S. (2013a). Reassortment between two serologically unrelated bluetongue virus strains is flexible and can involve any genome segment. *Journal of Virology* 87: 543-557
- Shaw, A.E., Ratinier, M., Nunes, S.F., Nomikou, K., Caporale, M., Golder, M., Allan, K., Hamers, C., Hudelet, P., Zientara, S. et al., (2013b). Reassortment between two serologically unrelated bluetongue virus strains is flexible and can involve any genome segment. *Journal of Virology* 87: 543-557
- Shirafuji, H., Yanase, T., Kato, T., Yamakawa, M. (2012). Genetic and phylogenetic characterization of genome segments 2 and 6 of bluetongue virus isolates in Japan from 1985 to 2008. *The Journal of General Virology* 93: 1465-1473
- Singh, K.P., Maan, S., Samuel, A.R., Rao, S., Meyer, A.J., Mertens, P.P.C. (2004). Phylogenetic analysis of bluetongue virus genome segment 6 (encoding VP5) from different serotypes. *Veterinaria Italiana* 40: 479-483
- Sreenivasulu, D., Rao, M.V.S., Gard, G. (1999). Isolation of bluetongue virus serotype 2 from native sheep in India. *The Veterinary Record* 144: 452-453
- Sung, P.Y., Roy, P. (2014). Sequential packaging of RNA genomic segments during the assembly of Bluetongue virus. *Nucleic Acids Res* 42: 13824-13838
- Tamura, K., Peterson, D., Peterson, N., Stecher, G., Nei, M., Kumar, S. (2011). MEGA5: molecular evolutionary genetics analysis using maximum likelihood, evolutionary distance, and maximum parsimony methods. *Molecular Biology and Evolution* 28: 2731-2739
- Ting, L.J., Lee, M.S., Huang, T.S., Huang, C.C., Kuo, S.T., Lee, F., Jong, M.H., Shiau, J.R., Lin, S.Y. (2005). Identification of bluetongue virus in goats in Taiwan. *The Veterinary Record* 156: 52
- Zerbino, D.R., Birney, E. (2008). Velvet: algorithms for de novo short read assembly using de Bruijn graphs. *Genome Research* 18: 821-829
- Zhu, J., Yang, H., Li, H., Xiao, L., Wang, J., Li, N., Zhang, N. (2013). Full-genome sequence of bluetongue virus serotype 1 (BTV-1) strain Y863, the first BTV-1 isolate of eastern origin found in China. *Genome announcements* 1: e00403-00413

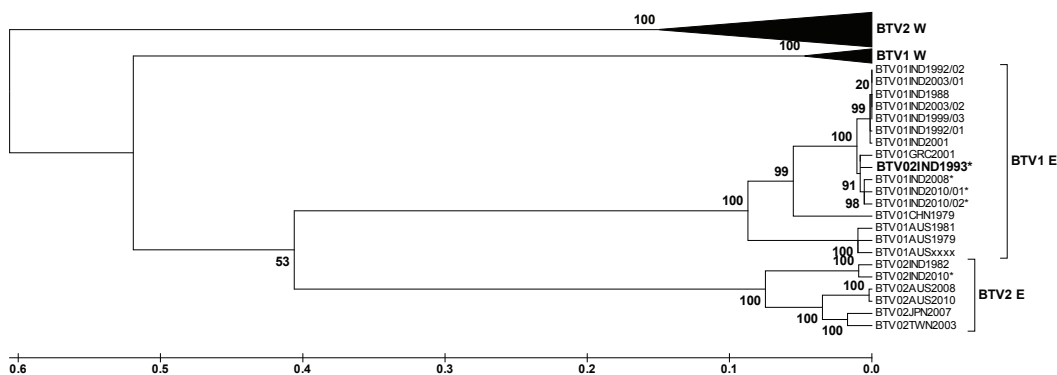


Fig 1: Phylogenetic analysis of segment 6 BTV serotypes 1 and 2.

Maximum Likelihood tree of Seg-6 of BTV serotype 1 and 2 using MEGA5.03 The percentage of trees in which the associated taxa clustered together is shown next to the branches. *Sequenced during this study

Table 1: Details of BTV sequences analysed

Virus ID	Serotype	Place of Isolation	Year of isolation	Acc. No	Reference
BTV01ZAF1958	BTV-1	South Africa	1958	JX272384	
BTV01AUS1979	BTV-1	Australia	1979	JN881990	(Boyle et al., 2012)
BTV01CHN1979	BTV-1	China	1979	KC879620	(Zhu et al., 2013)
BTV01AUS1981	BTV-1	Australia	1981	AJ631214	
BTV01NGA1982	BTV-1	Nigeria	1982	AJ586657	(Singh et al., 2004)
BTV01SDN1987	BTV-1	Sudan	1987	AJ586658	(Singh et al., 2004)
BTV01IND1988	BTV-1	India	1988	AJ586661	(Singh et al., 2004)
BTV01IND1992/01	BTV-1	India	1992	AJ586659	(Shaw et al., 2013b)
BTV01IND1992/02	BTV-1	India	1992	AJ586660	(Singh et al., 2004)
BTV01IND1999	BTV-1	India	1999	AJ586662	(Singh et al., 2004)
BTV01GRC2001	BTV-1	Greece	2001	AJ586664	(Singh et al., 2004)
BTV01IND2001	BTV-1	India	2001	AJ586663	(Singh et al., 2004)
BTV01IND2003/01	BTV-1	India	2003	AJ783903	
BTV01IND2003/02	BTV-1	India	2003	AJ783902	
BTV01DZA2006	BTV-1	Algeria	2006	EU422952	(Maan et al., 2008)
BTV01MAR2006	BTV-1	Morocco	2006	EU422953	(Maan et al., 2008)
BTV01FRA2007	BTV-1	France	2007	JX861493	(Shaw et al., 2013b)

BTV01IND2008*	BTV-1	Khammam	2008	JX399153	(Rao et al., 2013)
BTV01IND2010/1*	BTV-1	Nellore	2010	KP339149	(Reddy et al., 2016)
BTV01IND2010/2*	BTV-1	Karimnagar	2010	KP339139	(Reddy et al., 2016)
BTV01ITA2013	BTV-1	Italy	2013	KJ019210	(Lorusso et al., 2014)
BTV01AUSxxxx	BTV-1	Australia	xxxx	M21845	(Gould & Pritchard, 1988)
BTV02ZAF1959	BTV-2	South Africa	1959	JX272604	
BTV02IND1982	BTV-2	India	1982	AJ586675	(Singh et al., 2004)
BTV02NGA1982	BTV-2	Nigeria	1982	AJ586667	(Singh et al., 2004)
BTV02USA1982/01	BTV-2	USA	1982	AY855277	(Mecham & Johnson, 2005)
BTV02USA1982/02	BTV-2	USA	1982	AY855278	(Mecham & Johnson, 2005)
BTV02SDN1985	BTV-2	Sudan	1985	AJ586666	(Singh et al., 2004)
BTV02IND1993*	BTV-2	Chittor	1993	KP339159	(Sreenivasulu et al., 1999)
BTV02USA1999	BTV-2	USA	1999	AY855279	(Mecham & Johnson, 2005)
BTV02ITA2000/01	BTV-2	Italy	2000	JN255867	(Caporale et al., 2011)
BTV02ITA2000/02	BTV-2	Italy	2000	JN255877	(Caporale et al., 2011)
BTV02TUN2000	BTV-2	Tunisia	2000	AJ586668	(Singh et al., 2004)
BTV02FRA2001	BTV-2	France : Corsica	2001	AJ586674	(Singh et al., 2004)
BTV02ITA2001	BTV-2	Italy	2001	AJ586672	(Singh et al., 2004)
BTV02ITA2002	BTV-2	Italy	2002	AJ586670	(Singh et al., 2004)
BTV02TWN2003	BTV-2	Taiwan	2003	AY493690	(Ting et al., 2005)
BTV02JPN2007	BTV-2	Japan	2007	AB686238	(Shirafuji et al., 2012)
BTV02AUS2008	BTV-2	Australia	2008	JQ086246	(Boyle et al., 2012)
BTV02IND2010*	BTV-2	Karimnagar	2010	KP339169	(Reddy et al., 2016)
BTV02AUS2010	BTV-2	Australia	2010	JQ240326	(Boyle et al., 2012)
BTV02USA2010	BTV-2	USA	2010	JQ822253	(Maclachlan et al., 2013)
BTV02FRAxxxx	BTV-2	France :Corsica	xxxx	AY129083	
BTV02ITAxxxx	BTV-2	Italy	xxxx	AY189951	

* Sequenced during the current study

DEVELOPMENT OF GHEE RESIDUE CANDY INCORPORATED WITH ORANGE PEEL

Ananthakumar.K.V¹ and B.Dhanalakshmi²

Department of Livestock Products Technology (Dairy Science), Madras Veterinary College
Tamil Nadu Veterinary and Animal Sciences University, Chennai - 600 007.

ABSTRACT

This study was carried out to develop a candy utilizing the by-products of juice and dairy industry viz. orange peel and the ghee residue. Ghee residue candy with incorporation of dried orange peel powder and aqueous extract at various levels of 5,10,15 per cent (T_1, T_2, T_3) were prepared. The per cent inhibition of Di-phenyl-picryl-hydrazyl free radical antioxidant activity of fresh orange peel, orange peel powder and aqueous extract were found to be 76.91 ± 0.13 , 68.84 ± 0.22 , 72.45 ± 0.21 respectively. Nutritional, textural and sensory qualities of the ghee residue candy incorporated with various levels of orange peel was carried out. On sensory evaluation by using 9-point hedonic scale T_2 was found to be the best among the various levels tried. Texture analysis of ghee residue candy T_1, T_2 and T_3 were carried out for the parameters viz. hardness and stickiness and it was found that there was a significant difference between various inclusion level of orange peel powder and aqueous extract. The nutritional quality of the ghee residue candy (T_1, T_2 and T_3) were analyzed and it was found that, as the percentage of inclusion of orange peel increased there was a corresponding increase in the nutrients except nitrogen free extract. The ghee residue candy incorporated with orange peel T_2 was found to possess a higher antioxidant activity 70.44 ± 0.10 compared to control 10.50 ± 0.21 .

Key words: Ghee residue candy, Orange peel, Antioxidant activity, Vitamin C, Aqueous extract.

INTRODUCTION

Candy is a confectionary sweet food prepared mainly from sugar and liquid glucose. Candy is consumed globally by all age group people and is popular, especially among children. There is a high demand in global market for value added candies of functional importance. A vast variety of candies are available in international market

and is accepted by consumers based on its flavor and color. Developing an innovative candy with enhanced nutrients may add a new variety of the candy to the existing array of candies available in global market.

Candy is a sweet food prepared from fruits or vegetables by impregnating them with sugar syrup followed by draining of excessive syrup and then drying the product

¹Corresponding author e-mail id. anantha112@yahoo.co.in

²Professor and Head, Dept. of LPT (Dairy Science), Madras Veterinary College, Chennai.

to a shelf stable state. Fruits and vegetables like apples, ginger, mangoes, guava, carrot and citrus peels have been used to prepare candies (Mehta and Bajaj, 1984).

The ghee residue, a by-product of ghee is rich in phospholipids, sulphhydryl compounds, poly unsaturated fatty acids (PUFA), lipid, non-lipid constituents, flavor concentrates like FFA, carbonyls, and lactones (Serunjogi *et al.*, 1998). Antioxidant property of ghee residue is due to the mixture of its constituents. Ghee residue can be used as a source of natural antioxidant for improving the shelf life of food products including dairy products.

Orange constitutes about 60 per cent of the total citrus production (Lucia and Calogera, 2008). Peel represents between 50 to 65 per cent of total weight of the fruits and is the primary by-product (Ashbell and Donahaye, 1984). Orange peels are rich in flavonones, powerful antioxidants that help to reduce oxidative damage and fight free radicals.

Orange peels are also loaded with natural histamine suppressing compounds. The vitamin C in orange peel is about 136

mg/100 g whereas in the pulp it is about 50 mg/100 g (Zamantha *et al.*, 2014). It also contains considerable amounts of calcium, copper, magnesium, vitamin A, folate, other B complex vitamins and dietary fiber. Citrus peels contain principal dietary fiber sources such as cellulose, hemicelluloses, lignin, pectin, gums and bioactive compounds.

The present study was undertaken to develop ghee residue candy by incorporating orange peel, to effectively utilize the nutrients in the orange peel.

MATERIALS AND METHODS

Ingredients for ghee residue candy

Ghee residue, Skim milk powder, Orange peel, Liquid glucose, Sugar, Cocoa powder and dark chocolate were used for this study.

Standardization and selection of ingredients for preparation of ghee residue candy incorporated with orange peel

The ghee residue candy incorporated with orange peel was standardized using three different level of orange peel.

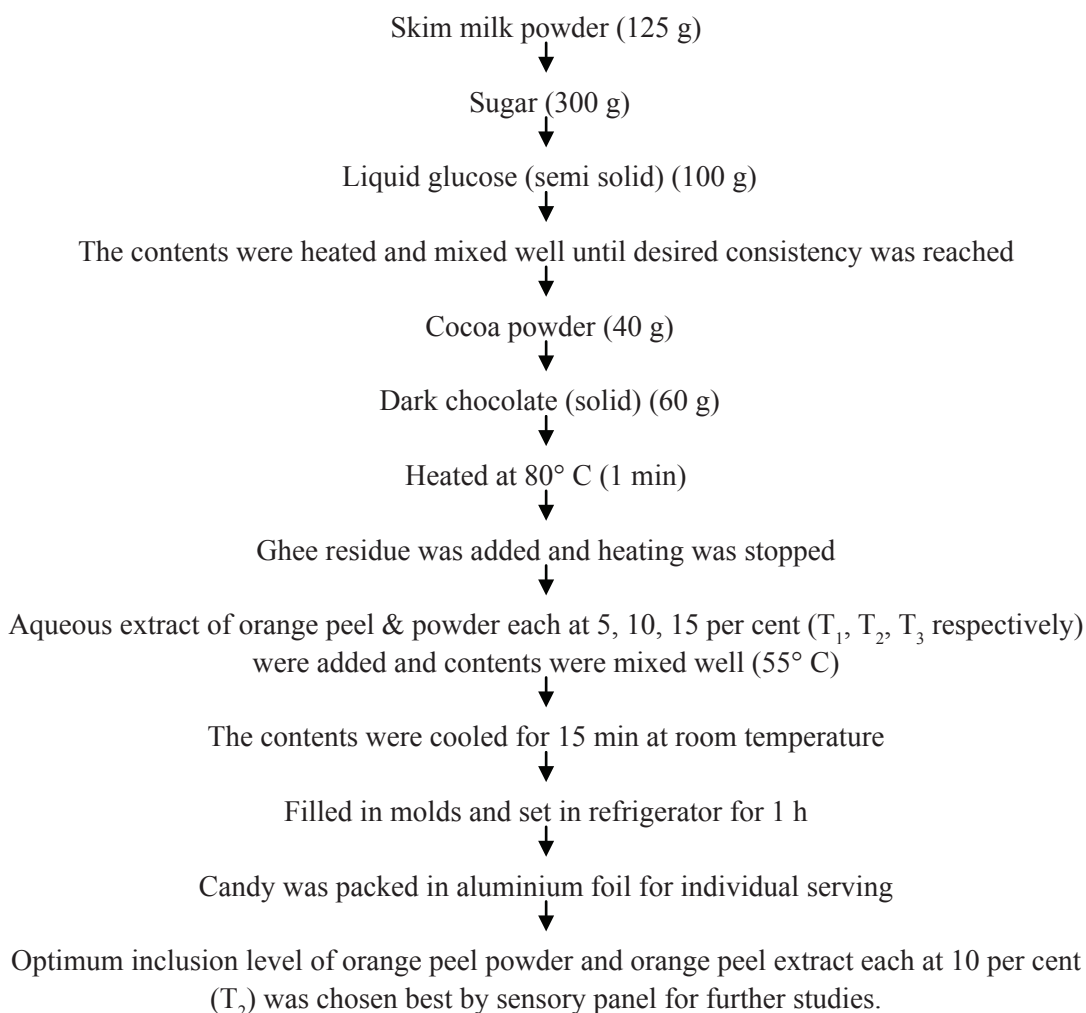
S.No.	Ingredients	Control	T ₁	T ₂	T ₃
1.	Skim milk powder	100g	100g	100g	100g
2.	Liquid glucose	80g	80 g	80g	80g
3.	Cocoa powder	32g	32g	32g	32g
4.	Dark chocolate (Morde 45 per cent cocoa)	48g	48g	48g	48g
5.	Ghee residue (processed)	---	500g	500g	500g
6.	Sugar	240g	240g	240g	240g
7.	Orange peel powder and aqueous extract	---	50g (5 per cent)	100g (10 per cent)	150g (15 per cent)

T₁ – Ghee residue candy incorporated with 5 per cent orange peel and extract.

T₂ - Ghee residue candy incorporated with 10 per cent orange peel and extract.

T₃ - Ghee residue candy incorporated with 15 per cent orange peel and extract.

**Flow chart for the preparation of ghee residue candy incorporated with orange peel
(modified technique of Wadhwa, 1997)**



Estimation of antioxidant activity

The antioxidant activity was assessed by DPPH free radical scavenging assay (Nur Alam *et al.*, 2013) to study the influence of added orange peel in the candy prepared using ghee residue.

Proximate analysis of ghee residue candy incorporated with orange peel (T₂)

The developed product was analyzed for moisture, crude protein, crude fiber, ether extract, nitrogen free extract and total ash as per the procedure described in AOAC (1997).

Sensory evaluation

Sensory evaluation was conducted by using the 9 point hedonic scale for the sensory attributes developed by Larmond (1997).

Texture analysis

The ghee residue candy incorporated with orange peel was observed for textural

properties, by using texture analyzer as per Afoakwa *et al.* (2007) with a penetration probe P/5 attached to a 50 kg load cell (Stable Micro Systems; Model : TA XT Plus), connected to a computer, programmed with texture analysis software, assembled for candy analysis.

RESULTS AND DISCUSSION

Table 1. Sensory evaluation of ghee residue candies incorporated with orange peel using 9-point hedonic scale (Mean + SE)[@]

Sensory attributes	Treatments				
	Control	T ₁	T ₂	T ₃	F-value
Colour	7.50±0.22	7.10±0.16	7.60± 0.30	7.33±0.21	2.312 ^{NS}
Flavour	7.15 ^d ±0.21	7.23 ^c ±0.30	8.00 ^a ± 0.22	7.30 ^b ±0.16	18.932 ^{**}
Texture	6.90 ^c ±0.19	7.30 ^a ±0.22	7.30 ^a ± 0.22	7.00 ^b ±0.21	5.098 ^{**}
Sweetness	6.67 ^d ±0.18	7.50 ^a ±0.22	7.00 ^b ±0.20	6.92 ^c ± 0.22	4.615 ^{**}
Overall acceptability	7.05 ^d ±0.25	7.28 ^b ±0.22	7.47 ^a ± 0.25	7.13 ^c ±0.16	5.942 ^{**}

[@]Average of six trials

Mean with different superscripts within a same row differ significantly from each other (P<0.01)

NS – Non significant (P > 0.05)

** Highly significant (P < 0.01)

T₁ – Ghee residue candy incorporated with 5 per cent orange peel and extract.

T₂ - Ghee residue candy incorporated with 10 per cent orange peel and extract.

T₃ - Ghee residue candy incorporated with 15 per cent orange peel and extract.

The sensory evaluation of ghee residue candy incorporated with orange peel at 5 per cent (T₁), 10 per cent (T₂) and 15 per cent (T₃) were conducted by using 9-point hedonic scale along with the control, revealed a highly significant difference (P < 0.01) for all sensory attributes viz. flavor, texture, sweetness and overall acceptability except for color. T₂ was found to be the best followed by T₁, T₃ and control. Highly significant difference

was noticed for the sensory attributes viz. texture, sweetness, overall acceptability between control and treatments. Negligible difference was noticed for texture between T₁ and T₂ whereas for sweetness T₁ scored higher scores. The decrease in sweetness might be due to the increase in addition of orange peel contributed by the higher fiber content in the orange peel as per Gorinstein *et al.* (2001). As per the scores for overall acceptability T₂ was found to be the best

followed by T₁, T₃ and control and hence of addition of orange peel in preparation of T₂ was considered to be the optimum level ghee residue candy.

Table 2. Nutritional composition of ghee residue candy incorporated with orange peel in per cent (Mean + SE) @

S.No.	Constituents	Treatments				F-value
		Control	T ₁	T ₂	T ₃	
1.	Moisture	5.26 ^c ± 0.22	10.46 ^b ± 0.12	10.60 ^b ± 0.11	11.45 ^a ± 0.11	343.388**
2.	Crude Protein	5.60 ^d ± 0.11	6.19 ^c ± 0.02	6.75 ^b ± 0.02	7.32 ^a ± 0.13	70.892**
3.	Crude Fibre	0.15 ^d ± 0.01	2.81 ^c ± 0.02	3.53 ^b ± 0.01	3.97 ^a ± 0.05	476.179**
4.	Ether extract	9.47 ^c ± 0.13	10.53 ^b ± 0.16	10.88 ^b ± 0.17	10.94 ^a ± 0.21	89.960**
5.	Total ash	0.99 ^d ± 0.04	1.09 ^c ± 0.15	1.49 ^b ± 0.04	1.72 ^a ± 0.04	50.764**
6.	Nitrogen free extract	78.53 ^a ± 0.32	68.92 ^b ± 0.17	66.75 ^c ± 0.17	64.60 ^d ± 0.34	516.647**

@Average of six trials

Mean with different superscripts within a same row differ significantly from each other (P<0.01)

** Highly significant (P < 0.01)

T₁ – Ghee residue candy incorporated with 5 per cent orange peel and extract.

T₂ - Ghee residue candy incorporated with 10 per cent orange peel and extract.

T₃ - Ghee residue candy incorporated with 15 per cent orange peel and extract.

The nutritional composition of control, 5 per cent (T₁), 10 per cent (T₂) and 15 per cent (T₃) ghee residue candy incorporated with orange peel revealed a highly significant difference (P < 0.01) for moisture, crude protein, crude fiber, ether extract, total ash and nitrogen free extract. All the nutrients were found to increase as level of inclusion of orange peel increased. Moisture content in ghee residue candy incorporated with orange peel when compared to control was found to increase when compared to control which might be due to the addition of aqueous extract of orange peel. Increase in crude protein per cent in treated sample might be due to the inclusion of ghee residue, cocoa powder

and orange peel in the candy as per Santha and Narayanan (1978) and Grohmann *et al.* (1995).

All the other nutrients like crude fiber, ether extract, total ash and NFE increased. This finding almost correlates with the findings of Nasser *et al.* (2008) and Magda *et al.* (2008) who stated that crude fiber, carbohydrates increased as the inclusion level of orange peel increased. These findings also correlate the findings of Hanan *et al.* (2012) who found that 10 per cent incorporation of citrus peel powders in wheat biscuits increased crude protein, crude fat contents as well as crude fiber and moisture content.

Table 3. Texture Analysis of ghee residue candy incorporated with orange peel (Mean + SE) @

Constituents	Control	Treatments			F – value
		T ₁	T ₂	T ₃	
Hardness (g)	100.60 ^d ± 0.36	102.58 ^a ± 0.31	101.38 ^b ± 0.34	101.36 ^b ± 0.20	9.433**
Stickiness (g)	-102.31 ^a ± 0.36	-102.04 ^b ± 0.38	-101.93 ^c ± 0.39	-101.44 ^d ± 0.40	4.616**

@Average of six trials

Mean with different superscripts within a same row differ significantly from each other (P<0.01)

** Highly significant (P < 0.01)

T₁ – Ghee residue candy incorporated with 5 per cent orange peel and extract.

T₂ - Ghee residue candy incorporated with 10 per cent orange peel and extract.

T₃ - Ghee residue candy incorporated with 15 per cent orange peel and extract.

The texture analysis of control and ghee residue candy incorporated with orange peel at 5 per cent (T₁), 10 per cent (T₂) and 15 per cent (T₃) were analyzed for the parameters hardness and stickiness revealed a highly significant difference (P < 0.01) between treatments.

Increase in hardness and decrease in stickiness in treated samples compared to control might be due to the hydrophilicity of orange peel in the ghee residue candy as per Nassar *et al.* (2008) and Kohajdova *et al.* (2011).

Table 4. Antioxidant activity of fresh orange peel, powdered dried orange peel and aqueous extract (Mean + SE) (per cent inhibition of DPPH free radical)@

Control (Ascorbic acid)	Fresh orange peel	Powdered dried orange peel	Aqueous extract	F – value
83.99 ^a ±0.21	76.91 ^b ± 0.13	68.84 ^d ± 0.22	72.45 ^c ±0.21	2346.9**

The antioxidant activity of fresh orange peel, powdered dried orange peel and aqueous extract of orange peel were 76.91±0.13, 68.84±0.22 and 72.45± 0.21 per cent inhibition of DPPH free radical respectively and a highly significant difference (P < 0.01) was noticed between them.

The decreased antioxidant activity exhibited in the powdered dried orange peel

might be due to the partial destruction of vitamin C which also possesses antioxidant property as per Lario *et al.* (2004). The higher antioxidant activity of aqueous extract compared to powdered dried orange peel might be due to the higher concentration of flavonoides in the aqueous extract which possess antioxidant activity as per Chakraborty and Shah (2011).

Table 5. Antioxidant activity of ghee residue candy incorporated with orange peel (T₂) (Mean + SE) (per cent inhibition of DPPH free radical)[@]

Antioxidant activity per cent inhibition of DPPH free radical		t - value
Control candy with Ghee residue	Ghee residue candy incorporated with orange peel (T ₂)	
10.50 ^b ± 0.21	70.44 ^a ±0.10	13435.9**

[@]Average of six trials

Mean with different superscripts within a same row differ significantly from each other (P<0.01)

** Highly significant (P < 0.01)

T₂ - Ghee residue candy incorporated with 10 per cent orange peel and extract.

Ghee residue candy incorporated with orange peel (T₂) and control candy were 10.50±0.21 and 70.44±0.10 per cent inhibition of DPPH free radical and revealed a highly significant difference (P < 0.01) between control and ghee residue candy.

The increase in antioxidant activity ghee residue candy incorporated with orange peel compared to control is due to the incorporation of orange peel which contains vitamin C and phenolic compounds which have the ability to scavenge the reactive oxygen capable of causing oxidative effect as per Gil- Izquierdo *et al.* (2001).

CONCLUSION

Candy, also known as sweet or lolly, is a confection that features sugar as a principal ingredient. Vegetables, fruit, or nuts which have been glazed and coated with sugar are said to be candied. Physically, candy is characterized by the use of a significant amount of sugar or sugar substitutes.

Sensory analysis revealed that ghee residue candy incorporated with orange peel (T₂) was preferred than other levels of inclusion. Increase in crude protein per cent in treated sample is due to the inclusion of ghee residue in the candy. Increase in hardness and decrease in stickiness in treated samples compared to control might be due to the hydrophilicity of orange peel in the ghee residue candy.

The antioxidant activity of the candy was carried out as ghee residue was used as one of the ingredients in the preparation of candy. The higher antioxidant activity of aqueous extract compared to powdered dried orange peel might be due to the higher concentration of flavonoides in the aqueous extract which possess antioxidant activity. The increase in antioxidant activity ghee residue candy incorporated with orange peel compared to control is due to the incorporation of orange peel which contains vitamin C and phenolic compounds which have the ability to scavenge the reactive oxygen capable of causing oxidative

effect. Hence the present has explored the possibilities to develop an innovative candy with enhanced nutrients to suite the palate of the consumers utilizing the by-products of dairy and juice industry.

REFERENCES

- Afoakwa, E.O., A. Paterson and M.Fowler, 2007. Factors influencing rheological and textural qualities in chocolate - A review. *Trends Food Sci. Technol.* **18**:290-298.
- AOAC, 1997. Official methods of analysis of AOAC international, 16thedn(Curniff, P.A. eds). *Virginia AOAC international*, pp. 25-28.
- Ashbell, G and E. Donahaye, 1984. Losses in orange peel silage. *J.Animal feed Sci. Technol.* **68**: pp.73-77.
- Chakraborty, D and B. Shah, 2011. Antimicrobial, anti-oxidative and anti-hemolytic activity of Piper betel leaf extracts. *Int. Pharma.Pharmaceutical Sci.* **3**(3): 193-199.
- Gil-Izquierdo, A., M.I. Gil, F. Ferreres and F.Tomas-Barberan, 2001. In vitro availability of flavanoids and other phenolics in orange juice. *J.Agric. Food Chem.* **49**: 1035–1041.
- Gorinstein, S., O. Martin-Belloso, Y. S. Park, R. Haruenkit, A. Lojek and M. Ciz, 2001. Comparison of some biochemical characteristics of different citrus fruits. *Food Chem.* **74**: 309–315.
- Grohmann, K., R.G. Cameron and B.S. Buslig, 1995. Fractionation and pretreatment of orange peel by dilute acid hydrolysis. *Bioresour.Technol.* **54**: 129-141.
- Hanan, M.E.K., J.K. Youssef and M.A. Rasha Mousa, 2012. Nutritional assessment of wheat biscuit and fortified wheat biscuits with Citrus Peel Powders. *J.Food Pub. Health.* **2**:55-60.
- Kohajdova, Z., J. Karovicova, M. Jurasovand and K. Kukurova, 2011. Application of citrus dietary fibre preparations in biscuit production. *J. Food and Nutr. Res.* **3**:182-190.
- Lario, Y., E. Sendra, , J. Garcí'a-Pe'rez, C. Fuentes, E. Sayas-Barberaand J. Ferná'ndez-Lo'pez, 2004. Preparation of high dietary fiber powder from lemon juice by-products.*Innov. Food Sci. Emerg. Technol.* **5**: 113–117.
- Larmond, E. 1997. Laboratory methods of sensory evaluation of foods. Canada. *Dept. Agric. Pub.No.1284.* pp. 36 – 37.
- Lucia, C and P. Calogera, 2008. Antioxidant activities of orange peel extracts. *J. World Appl. Sci.* **18**(5): 684-688.
- Magda, R.A., A.M. Awad and K.A. Selim, 2008. Evaluation of mandarin and navel orange peels as natural sources of antioxidant in biscuits. *Alex. J. Food Sci. Technol.* pp. 75-82.
- Mehta. U and S. Bajaj, 1984. Changes in the chemical composition and organoleptic quality of citrus peel

- candy during preparation and storage. *J Food Sci. Technol.* **21**:422–424.
- Nassar, A.G, A.A. Abdel-Hamied and E.A. El-Naggar 2008. Effect of citrus by-products flour incorporation on chemical, rheological and organolepic characteristics of biscuits. *World J. Agric. Sci.* **4**(5): 612-616.
- Nur Alam., J.B. Nusrat and Md. Rafiquzzaman, 2013. Review on in vivo and in vitro methods evaluation of antioxidant activity. *Saudi Pharma.J.* **21**:143-152.
- Santha, I.M and K.M. Narayanan, 1978. Chemical composition of ghee residue. *J.Food Sci. Tech.* **15**(1):24-27.
- Serunjogi. L., R.K. Abrahamsen and J. Narvhus, 1998. A Review paper: Current Knowledge of ghee and related products. *Int. Dairy. J.* **8**:677-688.
- Wadhwa, B.K.1997. Functional properties of ghee residue. “Technological advances in dairy by-products”, NDRI Publications. pp.141-146.
- Zamantha E. A., G.U. Janet, V.F. Aurora, J.T. Antonio and W.C. Jorge, 2014. Phytochemicals and antioxidant activity of juice, flavedo, albedo and comminuted orange. *J.Funct.foods.* **6**:470-481.

INFLUENCE OF HOUSING SYSTEM ON CUMULATIVE FEED CONSUMPTION AND FEED CONVERSION RATIO OF BROILERS FED WITH DIFFERENT LEVELS OF ENERGY AND PROTEIN

**K. Rajendran*, K. Mani, P. Shamsudeen,
V. Ramesh Saravana Kumar and A. Natarajan**

Department of Poultry Science, Veterinary College and Research Institute,
Namakkal- 637 002. Tamil Nadu

ABSTRACT

Commercial broilers were fed with different levels of energy (2850, 2950 and 3050 kcal/kg in pre-starter diet, 2950, 3050 and 3150 kcal/kg in starter diet and 3050, 3150 and 3250 kcal/kg in finisher diet) and protein (21.5, 22.5 and 23.5% in pre-starter diet, 20.5, 21.5 and 22.5% in starter diet and 19, 20 and 21% in finisher diet) for a period of five weeks to assess the production performance in both environmentally controlled and open curtained housing systems. All the chicks were reared up to five weeks under standard managerial condition. During this experimental period, data on body weight, feed consumption and mortality if any were recorded at weekly interval. The result of the study revealed lower feed intake in group T₇ (2640.16 g) in environmentally controlled housing system and group T₆ (2667.19 g) in open curtained housing system. The broilers reared in environmentally controlled housing system had significantly better feed conversion ratio (1.31 to 1.42) than broilers reared in open curtained housing system (1.45 to 1.52).

Key words: Housing system, Cumulative feed consumption, Feed conversion ratio, Broilers.

INTRODUCTION

Indian broiler industry has gone through tremendous development and expansion during the last two decades. Increased genetic potential of broilers coupled with higher placements helped in rapid development of broiler production in hot climates and requires greater emphasis on finding solutions to alleviate growth depression due to heat stress. Ambient temperature is an important determinant of bird performance. The main consequence of

heat exposure is reduction in feed intake in order to reduce metabolic heat production. In broilers this reduction is approximately 1.5 to 2.5 per cent per °C increase in ambient temperature above 20°C. Reduction in growth is often greater than the reduction in feed intake, resulting in lower feed efficiency.

Prevailing conditions of increasing ambient temperature and system of rearing in tropical countries are a major concern in broiler production. The future of broiler

*Corresponding author: Email: poultryrajendran@yahoo.com

industry may increasingly depend on environmentally controlled poultry house rather than open curtained poultry house, due to conditions like global warming. In India, studies are being focused primarily on the requirement of energy and protein and their proportion for augmentation of broiler growth only for open sided houses.

MATERIALS AND METHODS

The biological experiments were designed and conducted during January to February 2014 in environmentally controlled broiler house as well as in open sided broiler house located at the Poultry Farm Complex, Department of Poultry Science, Veterinary College and Research Institute, Namakkal, Tamil Nadu. Namakkal is situated in northwestern agro-climatic zone of Tamil Nadu at 11°2' N latitude and 78°2' E longitude at an altitude of 192 m above the mean sea level. The minimum and maximum temperature and relative humidity during the study period were 19.82 °C and 30.43 °C and 52.12 and 77.45 per cent, respectively.

The biological experiment was carried out with five hundred and seventy six (each 288 in environmentally controlled deep litter house and open sided deep litter house), sex separated, day-old, commercial (Vencobb 400) broiler chicks belonging to single hatch purchased from local hatchery.

Chicks were wing banded, weighed and randomly allotted into nine treatment groups with four replicates of eight chicks in each under both system of rearing.

Throughout the study period of five weeks, data on body weight, feed intake, mortality if any were recorded at weekly interval. The data collected on feed intake, body weight and calculated data on feed conversion ratio were subjected to statistical analysis as per the method suggested by Snedacor and Cochran (1989). Angular transformation was applied to percentages wherever needed before carrying out statistical analysis. The treatments applied to the nine groups of the experiment are provided in Table I.

Table-I

Treatment groups for each system of rearing	Particulars			Number of replicates per treatment	Number of birds per replicate	Total number of birds per treatment
	Type of feed	CP (%)	ME (kcal/kg)			
T ₁	Pre-starter	21.5	2850	4	8	32
	Starter	20.5	2950			
	Finisher	19.0	3050			
T ₂	Pre-starter	22.5	2850	4	8	32
	Starter	21.5	2950			
	Finisher	20.0	3050			

T ₃	Pre-starter	23.5	2850	4	8	32
	Starter	22.5	2950			
	Finisher	21.0	3050			
T ₄	Pre-starter	21.5	2950	4	8	32
	Starter	20.5	3050			
	Finisher	19.0	3150			
T ₅	Pre-starter	22.5	2950	4	8	32
	Starter	21.5	3050			
	Finisher	20.0	3150			
T ₆	Pre-starter	23.5	2950	4	8	32
	Starter	22.5	3050			
	Finisher	21.0	3150			
T ₇	Pre-starter	21.5	3050	4	8	32
	Starter	20.5	3150			
	Finisher	19.0	3250			
T ₈	Pre-starter	22.5	3050	4	8	32
	Starter	21.5	3150			
	Finisher	20.0	3250			
T ₉	Pre-starter	23.5	3050	4	8	32
	Starter	22.5	3150			
	Finisher	21.0	3250			
Total						288

RESULTS AND DISCUSSION

Effect of housing system on mean (\pm S.E.) feed consumption (g/bird) of broilers from first to fifth week of age as influenced by different levels of energy and protein is given in Table III.

The results revealed that at first week of age, broilers reared in environmentally controlled housing system recorded higher feed consumption than the broilers reared in open sided housing system. At the end of fifth week, the feed consumption of broilers

reared in environmentally controlled housing system was significantly higher than broilers reared in open sided housing system in all treatment groups indicating that housing system influenced cumulative feed consumption in broilers, irrespective of various combinations of dietary energy and protein.

The result of the study revealed a lower feed intake was noticed in group T₇ (2640.16 g) in environmentally controlled housing system and group T₆ (2667.19 g) in open sided housing system.

The group T₇ was fed with broiler pre-starter diet with 3050 kcal/kg of ME and 21.5% CP, broiler starter diet with 3150 kcal/kg of ME and 20.5% CP and broiler finisher diet with 3250 kcal/kg of ME and 19% CP. The group T₆ was fed with broiler pre-starter diet with 2950 kcal/kg of ME and 23.5% CP, broiler starter diet with 3050 kcal/kg of ME and 22.5% CP and broiler finisher diet with 3150 kcal/kg of ME and 21% CP whereas highest feed intake was recorded in group T₃ reared in both the housing system. The group T₃ was fed with broiler pre-starter diet with 2850 kcal/kg of ME and 23.5% CP, broiler starter diet with 2950 kcal/kg of ME and 22.5% CP and broiler finisher diet with 3050 kcal/kg of ME and 21% CP.

Effect of housing system on feed conversion ratio of broilers from first to fifth week of age as influenced by different levels of energy and protein is given in Table IV.

The broilers reared in environmentally controlled housing system had significantly better feed conversion ratio (1.38 to 1.52) than the broilers reared in open sided housing system (1.47 to 1.68). Interaction of dietary treatments and housing system recorded significant and highly significant difference between treatment groups at fifth week feed conversion ratio in broilers. This indicated that the housing system influenced the feed conversion ratio irrespective of various combinations of dietary energy and protein in broilers.

Within the same dietary protein and energy level, environmentally controlled housing system recorded approximately 10 per cent better feed conversion ratio than open sided housing system.

The result of the study indicated that the cumulative feed conversion ratio was best in group T₇ for broilers in environmentally controlled housing system. The group T₇ was fed with broiler pre-starter diet with 3050 kcal/kg of ME and 21.5% CP, broiler starter diet with 3150 kcal/kg of ME and 20.5% CP and broiler finisher diet with 3250 kcal/kg of ME and 19% CP. In case of open sided housing system group T₆ showed better result which was fed with broiler pre-starter diet with 2950 kcal/kg of ME and 23.5% CP, broiler starter diet with 3050 kcal/kg of ME and 22.5% CP and broiler finisher diet with 3150 kcal/kg of ME and 21% CP.

The results are in accordance with the earlier works of Al-Batshan and Hussein (1999), Ghaffari *et al.* (2007b), Kamran *et al.* (2008a), Kabir *et al.* (2010), Moosavi *et al.* (2011) and Elagib and Elzubeir (2012) who also reported that broilers fed with diet containing high energy had less feed consumption.

Al-Batshan and Hussein (1999), Maiorka *et al.* (2005), Ghaffari *et al.* (2007a), Kabir *et al.* (2010), Jafarnejad and Sadegh (2011), Moosavi *et al.* (2011), Malomo *et al.* (2013) also reported similar results, that broilers fed with diet containing high energy showed better feed conversion.

Table II. Effect of housing system on mean (\pm S. E.) body weight (g) of broilers from 1 to 5 weeks of age as influenced by different levels of energy and protein

Treatment groups	House	I Week	II Week	III Week	IV Week	V Week
T ₁	OPHS	157.47 \pm 2.2	377.69 \pm 7.37	713.66 \pm 20.40	1212.22 \pm 18.96	1696.13 \pm 26.01
	EC	165.63 \pm 2.47	415.91 \pm 8.45	801.41 \pm 20.95	1317.13 \pm 22.87	1878.28 \pm 36.30
	"t" value	2.46*	3.40*	3.00*	3.53*	4.07**
T ₂	OPHS	158.46 \pm 2.69	390.70 \pm 6.77	788.75 \pm 10.03	1271.63 \pm 32.34	1772.82 \pm 18.91
	EC	158.00 \pm 1.73	413.41 \pm 6.34	815.66 \pm 11.51	1316.97 \pm 11.59	1856.22 \pm 36.00
	"t" value	1.37^{NS}	2.44^{NS}	3.71*	1.31*	2.05*
T ₃	OPHS	161.37 \pm 3.72	401.06 \pm 6.66	752.85 \pm 9.37	1289.15 \pm 23.76	1730.47 \pm 9.49
	EC	161.62 \pm 1.35	432.41 \pm 5.66	859.22 \pm 10.35	1382.06 \pm 18.37	1930.44 \pm 30.74
	"t" value	0.63^{NS}	3.58*	7.61*	3.07*	6.21**
T ₄	OPHS	164.03 \pm 1.16	389.37 \pm 10.57	762.75 \pm 13.64	1242.68 \pm 25.60	1720.59 \pm 28.45
	EC	162.09 \pm 1.73	428.91 \pm 5.97	840.68 \pm 6.11	1376.69 \pm 17.83	1951.44 \pm 20.54
	"t" value	9.28^{NS}	3.26*	5.21**	4.29**	6.57**
T ₅	OPHS	165.06 \pm 2.53	399.03 \pm 6.72	787.41 \pm 12.53	1300.35 \pm 10.04	1803.82 \pm 23.03
	EC	157.94 \pm 0.77	428.28 \pm 3.72	839.82 \pm 5.56	1369.66 \pm 17.85	1908.16 \pm 13.43
	"t" value	2.69*	3.86*	3.82*	3.38*	3.91*
T ₆	OPHS	168.00 \pm 2.78	400.60 \pm 8.93	790.63 \pm 15.76	1308.60 \pm 35.43	1819.25 \pm 29.16
	EC	159.75 \pm 3.99	426.16 \pm 8.94	855.25 \pm 13.07	1368.01 \pm 24.07	1937.79 \pm 57.06
	"t" value	1.69^{NS}	2.02*	3.15*	1.38*	1.85*
T ₇	OPHS	162.58 \pm 2.36	390.84 \pm 13.10	760.10 \pm 25.80	1261.45 \pm 27.78	1724.84 \pm 42.45
	EC	162.63 \pm 2.76	431.02 \pm 10.43	852.56 \pm 13.06	1377.74 \pm 13.56	1963.13 \pm 50.13
	"t" value	0.01^{NS}	2.40^{NS}	3.19*	3.76*	3.62*
T ₈	OPHS	162.63 \pm 3.61	395.13 \pm 8.82	773.19 \pm 14.90	1286.13 \pm 16.96	1757.63 \pm 44.48
	EC	160.57 \pm 1.61	426.97 \pm 6.00	852.35 \pm 19.90	1411.38 \pm 21.63	1989.60 \pm 47.68
	"t" value	0.52^{NS}	2.98*	3.18*	4.55**	3.57*
T ₉	OPHS	165.41 \pm 3.10	406.19 \pm 12.22	811.35 \pm 17.10	1372.10 \pm 25.45	1884.38 \pm 50.73
	EC	158.19 \pm 3.72	426.60 \pm 8.35	875.07 \pm 11.80	1448.47 \pm 17.60	2022.19 \pm 42.75
	"t" value	1.49^{NS}	1.37^{NS}	3.06*	2.46*	2.07*

OPHS - Open sided housing system, EC- Environmentally controlled housing system.

NS- Non significant, * - Significant (P<0.05) and ** - Highly significant (P<0.01)

Table-III. Effect of housing system on mean (\pm S. E.) cumulative feed consumption (g/bird) of broilers from 1 to 5 weeks of age as influenced by different levels of energy and protein

Treatment groups	House	I Week	II Week	III Week	IV Week	V Week
T ₁	OPHS	152.19 \pm 2.21	484.85 \pm 5.84	1009.85 \pm 25.65	1852.35 \pm 32.58	2846.25 \pm 12.95
	EC	145.60 \pm 1.54	348.32 \pm 7.12	1018.91 \pm 23.58	1861.41 \pm 31.54	2855.32 \pm 14.01
	"t" value	2.28^{NS}	7.12^{**}	11.24^{**}	7.12^{**}	3.37^{**}
T ₂	OPHS	150.38 \pm 2.95	481.03 \pm 6.25	990.10 \pm 12.05	1818.25 \pm 14.22	2802.28 \pm 28.22
	EC	138.13 \pm 2.56	348.13 \pm 5.01	995.31 \pm 13.51	1824.07 \pm 9.85	2807.50 \pm 30.14
	"t" value	2.31[*]	1.12^{NS}	4.66^{**}	5.17^{**}	2.95[*]
T ₃	OPHS	151.41 \pm 1.45	485.32 \pm 4.86	1021.88 \pm 14.57	1908.91 \pm 25.21	2860.00 \pm 35.21
	EC	145.16 \pm 2.01	346.88 \pm 3.54	1028.60 \pm 13.22	1815.63 \pm 23.35	2866.77 \pm 34.66
	"t" value	1.73^{NS}	4.76^{**}	6.06^{**}	6.54^{**}	3.06^{**}
T ₄	OPHS	146.88 \pm 1.89	470.00 \pm 11.26	959.69 \pm 25.25	1802.04 \pm 20.55	2703.60 \pm 18.96
	EC	140.63 \pm 1.02	333.13 \pm 12.54	963.44 \pm 28.69	1830.78 \pm 18.82	2771.41 \pm 12.45
	"t" value	2.98[*]	0.73^{NS}	4.48^{**}	3.86^{**}	2.47[*]
T ₅	OPHS	143.38 \pm 1.87	481.63 \pm 5.20	1027.25 \pm 9.20	1854.13 \pm 21.20	2759.60 \pm 35.21
	EC	137.13 \pm 1.20	350.69 \pm 4.56	1033.44 \pm 8.55	1797.82 \pm 18.98	2703.28 \pm 30.28
	"t" value	3.16[*]	1.83^{NS}	6.18^{**}	1.78^{NS}	1.18^{NS}
T ₆	OPHS	149.53 \pm 1.54	466.72 \pm 6.87	946.25 \pm 19.65	1745.63 \pm 25.65	2667.19 \pm 40.12
	EC	143.28 \pm 1.46	332.03 \pm 6.01	954.85 \pm 17.21	1754.22 \pm 24.55	2675.88 \pm 36.58
	"t" value	4.71^{**}	2.05^{NS}	1.61^{NS}	1.17^{NS}	0.74^{NS}
T ₇	OPHS	152.66 \pm 1.21	475.32 \pm 7.85	986.57 \pm 13.41	1785.94 \pm 18.54	2728.91 \pm 30.55
	EC	143.13 \pm 1.89	326.10 \pm 9.84	980.47 \pm 14.84	1726.72 \pm 21.70	2640.16 \pm 17.84
	"t" value	4.16^{**}	2.30[*]	4.86^{**}	4.06^{**}	4.49^{**}
T ₈	OPHS	148.60 \pm 8.54	481.57 \pm 12.88	981.88 \pm 34.58	1824.07 \pm 38.54	2732.82 \pm 48.86
	EC	142.35 \pm 5.86	352.66 \pm 9.54	995.32 \pm 33.85	1837.50 \pm 39.56	2746.25 \pm 45.62
	"t" value	3.83^{**}	0.92^{NS}	5.60^{**}	2.79[*]	2.38[*]
T ₉	OPHS	165.63 \pm 8.54	506.41 \pm 22.14	1008.28 \pm 20.65	1867.35 \pm 34.21	2808.91 \pm 42.10
	EC	137.50 \pm 2.10	350.63 \pm 10.35	990.00 \pm 16.32	1874.06 \pm 33.22	2780.31 \pm 40.65
	"t" value	2.88[*]	1.09^{NS}	2.81[*]	1.95^{NS}	0.91^{NS}

OPHS - Open sided housing system, EC- Environmentally controlled housing system.

NS- Non significant, * - Significant (P<0.05) and ** - Highly significant (P<0.01)

Table-IV. Effect of housing system on mean (\pm S. E.) cumulative feed conversion ratio of broilers from 1 to 5 weeks of age as influenced by different levels of energy and protein

Treatment groups	House	I Week	II Week	III Week	IV Week	V Week
T ₁	OPHS	0.97 \pm 0.03	1.29 \pm 0.04	1.42 \pm 0.07	1.53 \pm 0.03	1.68 \pm 0.03
	EC	0.88 \pm 0.01	1.19 \pm 0.01	1.27 \pm 0.01	1.41 \pm 0.01	1.52 \pm 0.02
	"t" value	3.34*	2.44*	2.17*	3.26*	4.29**
T ₂	OPHS	0.95 \pm 0.02	1.23 \pm 0.03	1.31 \pm 0.03	1.43 \pm 0.04	1.58 \pm 0.02
	EC	0.87 \pm 0.02	1.18 \pm 0.01	1.22 \pm 0.01	1.39 \pm 0.02	1.51 \pm 0.03
	"t" value	3.50*	1.93^{NS}	2.89*	1.10^{NS}	1.84^{NS}
T ₃	OPHS	0.94 \pm 0.02	1.21 \pm 0.03	1.36 \pm 0.03	1.48 \pm 0.03	1.65 \pm 0.02
	EC	0.90 \pm 0.01	1.14 \pm 0.02	1.20 \pm 0.01	1.39 \pm 0.01	1.49 \pm 0.01
	"t" value	1.92^{NS}	2.19*	5.55**	2.98*	7.77**
T ₄	OPHS	0.90 \pm 0.01	1.21 \pm 0.02	1.26 \pm 0.03	1.46 \pm 0.03	1.57 \pm 0.03
	EC	0.87 \pm 0.01	1.11 \pm 0.03	1.15 \pm 0.04	1.33 \pm 0.02	1.42 \pm 0.02
	"t" value	2.32*	2.82*	2.40*	3.33*	4.69**
T ₅	OPHS	0.87 \pm 0.01	1.21 \pm 0.01	1.31 \pm 0.01	1.43 \pm 0.02	1.53 \pm 0.01
	EC	0.87 \pm 0.01	1.14 \pm 0.01	1.23 \pm 0.01	1.32 \pm 0.02	1.42 \pm 0.02
	"t" value	0.18^{NS}	4.70*	4.39*	4.76**	5.81**
T ₆	OPHS	0.89 \pm 0.01	1.17 \pm 0.03	1.20 \pm 0.03	1.34 \pm 0.06	1.47 \pm 0.05
	EC	0.90 \pm 0.02	1.12 \pm 0.03	1.12 \pm 0.03	1.28 \pm 0.02	1.38 \pm 0.03
	"t" value	0.34^{NS}	1.17^{NS}	1.97*	0.85*	1.58*
T ₇	OPHS	0.94 \pm 0.01	1.22 \pm 0.06	1.30 \pm 0.06	1.42 \pm 0.04	1.59 \pm 0.05
	EC	0.88 \pm 0.02	1.09 \pm 0.04	1.15 \pm 0.03	1.26 \pm 0.01	1.37 \pm 0.03
	"t" value	2.20*	1.90^{NS}	2.38*	3.74*	3.65*
T ₈	OPHS	0.92 \pm 0.03	1.22 \pm 0.04	1.27 \pm 0.04	1.42 \pm 0.02	1.56 \pm 0.04
	EC	0.89 \pm 0.02	1.16 \pm 0.03	1.17 \pm 0.03	1.30 \pm 0.03	1.38 \pm 0.02
	"t" value	0.82^{NS}	1.24^{NS}	2.01*	3.72*	4.29*
T ₉	OPHS	1.00 \pm 0.06	1.25 \pm 0.05	1.25 \pm 0.04	1.36 \pm 0.04	1.50 \pm 0.05
	EC	0.87 \pm 0.02	1.15 \pm 0.02	1.13 \pm 0.01	1.30 \pm 0.01	1.38 \pm 0.01
	"t" value	2.02^{NS}	1.96^{NS}	2.56*	1.59^{NS}	2.24*

OPHS - Open sided housing system, EC- Environmentally controlled housing system.

NS- Non significant, * - Significant (P<0.05) and ** - Highly significant (P<0.01)

REFERENCES

- Al-Batshan, H.A. and E.O.S. Hussein, 1999. Performance and carcass composition of broilers under heat stress: The effects of dietary energy and protein. *Asian-Aust. Journal of Animal Sciences*, **12(6)**: 914-922.
- Elagib, H.A.A. and E.A. Elzubeir, 2012. The effect of feeding different levels of methionine and two levels of energy on broiler performance during heat stress. *Pak. J. Nut.*, **11**: 739-742.
- Ghaffari, M., M. Shivazad and M. Zaghari, 2007a. Effect of energy levels of diets formulated on total and digestible amino acid basis on performance of female broiler. *Pak. J. Bio. Sci.*, **10(23)**: 4323-4326.
- Jafarnejad, S. and M. Sadegh, 2011. The effects of different levels of dietary protein, energy and using fat on the performance of broiler chicks at the end of the third weeks. *Asian Journal of Poultry Science*. **5(1)**: 35-40.
- Kabir, M.A., K.M.S. Islam, M.R. Debi, S.K. Das and M. Biswas, 2010. Effect of different levels of energy and protein diets on productivity, meat quality and production cost of broilers. *International Journal of Biological Research*. **2(11)**: 1-5.
- Kamran, Z., M. Sarwar, M. Nisa, M.A. Nadeem, S. Mahmood, M.E. Babar and S. Ahmed, 2008. Effect of low protein diets having constant energy to protein ratio on performance and carcass characteristics of broiler chickens from one to thirty-five days of age. *Poultry Science*. **87**: 468-474.
- Maiorka, A., F. Dahlke, A.M. Penz and A.M. Kessler, 2005. Diets formulated on total or digestible amino acid basis with different energy levels and physical form on broiler performance. *Braz. J. Poult. Sci.*, **7**: 47-50.
- Malomo, G.A., S.A. Bolu and S.G. Olutade, 2013. Effects of dietary crude protein on performance and nitrogen economy of broilers. *Sustainable Agri. Res.*, **2(3)**: 52-54.
- Moosavi, M., M. Eslami, M. Chaji and M. Boujarpour, 2011. Economic value of diets with different levels of energy and protein with constant ratio on broiler chickens. *J. Anim. Vet. Adv.*, **10**: 709-711.
- Snedacor, G.W. and W.G. Cochran, 1989. Statistical methods, (8th edn.) Iowa state University Press, Ames, Iowa. Pp.xx+503.

ANTIMICROBIAL EFFECT OF *PSIDIUM GUAJAVA*, *MANGIFERA INDICA* AND *PUNICA GRANATUM* EXTRACTS AGAINST BOVINE MASTITIS PATHOGEN

Silambarasan D, Amsaveni S, Kalaiselvi L* and Ramesh S

Department of Veterinary Pharmacology and Toxicology

Madras Veterinary College, Chennai-600 007

ABSTRACT

The present study was carried out to assess the antibacterial activity of aqueous extracts of *Psidium guajava* leaves, *Mangifera indica* kernel and *Punica granatum* pericarp. Antimicrobial activity against *Escherichia coli*, *Staphylococcus aureus* and *Streptococcus* spp. isolated from bovine mastitis cases were tested by measuring MIC by Resazurin dye broth microdilution method. All the three plant extracts exhibited antibacterial activity against organism tested. Of the three extracts, *Punica granatum* extracts was found to have lower MIC of 2.5 mg/ml against *E. coli* and *Streptococcus* sp compared to other two extracts. However, *Mangifera indica* kernel extract was found to have good antibacterial activity against *Staphylococcus aureus* with a lower MIC of 0.625 mg/ml compared to other two extracts. The results of the present study confirmed the antibacterial potential of plant extracts. Further studies are required to explore the bioactive molecules which would probably become an alternative source of new and natural antibacterial agents.

Key Words: Mastitis - Antibacterial activity - *Psidium guajava*- *Mangifera indica* – *Punica granatum*.

INTRODUCTION

Bovine mastitis continues to be one of the most problematic diseases of the dairy cattle and it is usually treated with antimicrobial agents. There has been increasing incidence of resistance development against antimicrobial drugs commonly employed in the treatment of mastitis. Development of multidrug resistance in bacterial pathogens has led to treatment failure and it necessitates search

for new alternative drugs. Plants have been used for medicinal purposes since time immemorial and the role of medicinal plants in the discovery of new drugs is overwhelming. In India, large numbers of plants have been used traditionally for treatment of various ailments. In the present study, we have selected three medicinal plants viz., *Psidium guajava* (Common name: Guava), *Mangifera indica* (Common name: Mango) and *Punica granatum* (Common name: Pomegranate), to evaluate

* Corresponding author: email: kalaiselvivet@gmail.com

their antimicrobial activity against bovine mastitis pathogens.

MATERIALS AND METHODS

Plant Collection

Guava leaves (*Psidium guajava*) were collected from Villupuram district of Tamilnadu, India. Pomegranate fruits (*Punica granatum*) and mango (*Mangifera indica*) were purchased from local market in Chennai, Tamilnadu, India.

Extract preparation

Pomegranate pericarp, mango seed kernel and guava leaves were shade dried and powdered using blender. The aqueous extract of each plant material was prepared by dissolving 20 g of powdered plant material in 100 ml of triple distilled water. This solution was thoroughly mixed using magnetic stirrer for 6 hrs and kept overnight without any disturbance. Following day, the solution was strained using muslin cloth and then filtered twice using Whatman filter paper No.1. The filtrate thus obtained was transferred to sterile petri dish and dried in hot air oven at 48–50°C overnight. Dried extract was scraped using sterile surgical blade, weighed, transferred to sterile glass vials and stored at -20°C.

Bacterial strains

A total of 20 bacterial isolates from bovine clinical/subclinical mastitis cases were studied. These isolates were collected, characterized and maintained by Department of Veterinary Pharmacology and Toxicology, MVC, Chennai. Isolates tested included *Staphylococcus aureus* (8

isolates), *Escherichia coli* (10 isolates) and *Streptococcus* spp (2 isolates).

Antimicrobial susceptibility testing

The MICs of plant extracts were determined by Resazurin dye broth microdilution method (Sarker *et al.*, 2007). Serial two fold dilutions of drugs in nutrient broth (50 µl) were prepared in 96 well ELISA plates. 10 µl of Resazurin dye, 30 µl of nutrient broth and 10 µl of bacterial suspension (adjusted to 5×10^6 CFU/ml) were added to the wells. Sterility control without bacteria and growth control without drug were also included. Plates were wrapped and incubated at 37°C for overnight. The lowest concentration that inhibited visible growth (persistence of blue color) of bacteria was recorded as MIC. The MICs at which 50% (MIC₅₀) and 90% (MIC₉₀) of the isolates were inhibited were calculated. All the experiments were performed in triplicate

RESULTS AND DISCUSSION

The MICs of the plant extracts against bacterial isolates are presented in Table 1. The aqueous extract of all the three plants showed antibacterial activity against both Gram positive (*Streptococci* spp. and *Staphylococcus aureus*) and Gram negative (*Escherichia coli*) organisms tested.

In the present study, it was observed that the extracts of mango seed kernel showed antibacterial activity against organisms tested. Similarly, Sowmiya *et al* (2009) reported that aqueous and ethanol extracts of mango kernel showed good antibacterial activity against *E. coli*, *S. aureus* and *S. pyogenes*. The extract of mango seed kernel

showed excellent activity against *S. aureus* with MIC of 0.3125-5 mg/l compared to other two extracts. Mirghani *et al.* (2009) reported that extracts of mango seed kernel are more effective against Gram-positive than the Gram-negative bacteria. In our study, similar result was found in which MIC of mango seed kernel against *E. coli* (MIC 5 mg/l) was higher than *S. aureus* (MIC₅₀ 0.625 mg/l). Antibacterial activity of mango seed kernel may be due to the presence of considerable amount of total phenolic compounds (Mirghani *et al.*, 2009). The presence of phellandrene, α -pinene, ambolic acid, ascorbic acid, β -carotene, gallic acid, gallotannic acid, mangifelic acid, mangiferol peroxidase, phenyl alanine and proline (Kabuki *et al.*, 2000) and gallotannins (Engels *et al.*, 2009) in mango seed kernel extracts showed antimicrobial activity to both Gram positive and Gram negative bacteria.

The extract of *Punica granatum* was equally effective in inhibiting both Gram positive and Gram negative organisms tested. These results are in agreement with those reported by Kadi *et al* (2011). They reported that extracts of *Punica granatum* bark showed broad spectrum of activity against both Gram positive and Gram negative organisms. We found that extract

of *Punica granatum* was more effective than the other two extracts against *E. coli* and *Streptococcus* spp. Silva *et al.* (2008) reported that extract of *Punica granatum* was effective against *S. aureus* isolated from bovine mastitis and can be used as an alternative therapy in veterinary medicine. The broad-spectrum antibacterial activity of this plant extract might be due to the presence of secondary metabolites such as tannins, alkaloids, phenolic compounds or saponins (Machado *et al.*, 2002).

We observed that guava leaf extract inhibited both Gram positive and Gram negative organisms with an MIC of 5.0 mg/ml. Ajariyakhajorn and Samnagamnim (2005) also reported that guava leaf extracts were effective against *S. aureus* isolates from bovine mastitis with a MIC of 0.2-51.2 mg/l. The active ingredients present in guava leaves include alkaloids, saponins, tannins, phenols, flavonoids, triterpenoids and steroids. (Arima and Danno, 2002)

This study reconfirmed the effectiveness of the extracts of *Punica granatum*, *Psidium guajava* and *Mangifera indica* as potential antibacterial agents against mastitis pathogens. Further studies are needed to elucidate the horizon of antibacterial activity and the structural information of the biomolecules.

Table 1 MIC results of plant extracts against bacterial isolates from bovine mastitis

Organism	Extract	MIC	MIC ₅₀
<i>Escherchia coli</i>	<i>Punica granatum</i> (mg/ml)	2.5	2.5
	<i>Mangifera indica</i> (mg/ml)	5.0	5.0
	<i>Psidium guajava</i> (mg/ml)	5.0	5.0
	Enrofloxacin (µg/ml)	2.44 – 156.25	2.44
<i>Staphylococcus aureus</i>	<i>Punica granatum</i> (mg/ml)	2.5	2.5
	<i>Mangifera indica</i> (mg/ml)	0.3125-5.0	0.625
	<i>Psidium guajava</i> (mg/ml)	5.0	5.0
	Enrofloxacin (µg/ml)	0.305 – 2.44	1.22
<i>Streptococcus</i> sp	<i>Punica granatum</i> (mg/ml)	2.5	2.5
	<i>Mangifera indica</i> (mg/ml)	2.5	2.5
	<i>Psidium guajava</i> (mg/ml)	5.0	5.0
	Ceftriaxone (µg/ml)	0.625	0.625

ACKNOWLEDEMENT

We thank the Dean, MVC, Chennai, for giving permission to carry out student Project in the Department of Veterinary Pharmacology and Toxicology, MVC, Chennai.

REFERENCES

- Ajariyakhajorn, K. and Samngamnim, S. (2005). Antibacterial activity of some herb extracts against bovine mastitis pathogens. The 4th Chulalongkorn University Veterinary Annual conference, Chulalongkorn University. p 102
- Arima, H. and Danno, G. (2002). Isolation of antimicrobial compounds from guava (*Psidium guajava* L.) and their structure elucidation. *Bioscience, Biotech. Biochem.*, **66(2)**: 1727-1730.
- Engels, C., Knodler, M., Zhao, Y.Y., Carle, R., Ganzle, M.G. and Schieber, A. (2009) Antimicrobial activity of gallotannins isolated from mango (*Mangifera indica* L.) kernels. *J Agr Food Chem.*, **57**: 7712-7718
- Kabuki, T., Nakajima, H., Arai, M., Ueda, S., Kuwabara, Y. and Dosako, S. (2000) Characterization of novel antimicrobial compounds from mango (*Mangifera indica* L.) kernel seeds. *Food Chemistry*, **71**: 61-66.
- Kadi, H., Moussaoui, A., Benmehdi, H., Lazouni, H.A., Benayahia, A. and Boudierba, N.N. (2011). Antibacterial activity of ethanolic and aqueous extracts of *Punica granatum* L. bark. *Journal of Applied Pharmaceutical Science*, **01 (10)**; 2011: 180-182

- Machado, T.B., Leal, I.C.R., Amaral, A.C.F., Santos, K.R.N., Silva, M.G., Kuster, R.M. 2002. Atimicrobial ellagitannin of *Punica granatum* fruits. *J Braz Chem Soc* **13(5)**: 606-610.
- Mirghani, M.E.S., Yosuf, F., Kabbashi, N.A., Vejayan, J. and Yosuf, Z.B.M. (2009). Antibacterial activity of mango kernel extracts. *J. Applied Sci.*, **9**: 3013-3019.
- Sarker, S.D., Nahar, L., Kumarasamy, Y. (2007). Microtitre plate-based antibacterial assay incorporating resazurin as an indicator of cell growth, and its application in the in vitro antibacterial screening of phytochemicals. *Methods*, **42**: 321-324.
- Silva, M.A.R., Higino, J.S., Pereira, J.V., Siqueira-Junior, J.P. and Pereira, M.S.V. (2008) Antibiotic activity of the extract of *Punica granatum* Linn over bovine strains of *Staphylococcus aureus*. *Rev. bras. Farmacogn*, **18 (2)**: 209-212
- Sowmiya, S., Soundarapandian, P. and Rajan, S. (2009). Bioactive Studies of *Mangifera indica* aganist Bacteria isolated from Urine Samples. *Current Research Journal of Biological Sciences*, **1(3)**: 139-143, 2009

EVALUATION OF POSTERIOR CAPSULAR OPACIFICATION IN DOGS – A REVIEW OF 12 CASES

Gokilavani. M¹, C. Ramani, Ph.D., DAiCVO², Mohamed Shafiuzaama Ph.D.³
S. Ramesh Ph.D.⁴

ABSTRACT

Posterior capsule opacification (PCO) is the most common long-term postoperative complication following cataract surgery. It causes impaired visual acuity by directly blocking the visual axis. A total of 12 cases were included in the study. The cases were divided into two groups, group I (cyclosporine A) and group II (mitomycin C) consisting of six eyes each. The concomitant preoperative conditions (stage of cataract, age of the patient, etc.) and intra operative conditions (Phacoemulsification parameters: power, irrigation aspiration fluid volume, time) were related with the development of post-operative complications. Evaluation and grading of PCO by using indirect ophthalmoscopy and EPCO 2000 software and the mean average of PCO scores in visual assessment by using indirect ophthalmoscopy was 0.29 ± 0.18 and mean PCO score in EPCO software score was 0.37 ± 0.1 . A higher PCO score were found on EPCO 2000 software than visual function assessment by using indirect ophthalmoscopy. This study was concluded that the software method of evaluation of PCO was easy, applicable and more appropriate method to evaluate in clinical studies.

Key words: PCO, Cyclosporine A, Mitomycin C

INTRODUCTION

Posterior capsule opacification (PCO) is the most common long-term postoperative complication following cataract surgery. It causes impaired visual acuity by directly blocking the visual axis.

Posterior capsule opacification (PCO) is the most common long-term postoperative complication following cataract surgery, also called as after cataract or secondary cataract. PCO results from the growth and abnormal proliferation of Lens Epithelial Cells (LECs) on the capsule at the time of cataract surgery. These cells migrate to the posterior capsule where they approach the central visual axis and cause visual axis obstruction, resulting in dimness

of vision⁽²⁾ The severity of PCO was greater in canine patients which might be related to the differences in inflammatory and fibrous response. While it was more difficult to quantify, it was likely that PCO represents a significant cause of reduced visual acuity in dogs after cataract surgery⁽¹⁾

MATERIALS AND METHODS

The present study was conducted at Small Animal Ophthalmology Unit of the Madras Veterinary College Teaching Hospital, over a period of 12 months. Clinical cases which were presented with a history of visual deficit and clinical signs suggestive of cataract, were included in this study. The patients were further screened for routine hematological (hemoglobin,

packed cell volume, RBC count) and serum biochemical (blood glucose level) parameters as an assessment of preoperative health status. Twelve cases were randomly selected after phacoemulsification surgery. In all 12 cases, the pupil dilatation was achieved using 0.8% tropicamide eye drops.

Two methods were used for the evaluation of PCO post operatively after phacoemulsification surgery. After pupil dilatation both indirect ophthalmoscopy and EPCO 2000 software⁽³⁾ evaluation were done in all the 12 cases.

RESULTS AND DISCUSSION

On EPCO software analysis, PCO could be well observed in 9 out of 12 cases. PCO after 50 days postoperative examination period. On visual assessment by using indirect ophthalmoscopy method PCO could be observed only in 5 out of 12 cases on 50th day postoperatively.

More number of PCO could be well observed and viewed in EPCO 2000 software method than visual assessment using indirect ophthalmoscopy method.

The mean of PCO score in visual assessment by using indirect ophthalmoscopy was **0.29 ± 0.18** and mean PCO score in EPCO software score was **0.37 ± 0.1**. A higher PCO score were found on EPCO 2000 software than visual

function assessment by using indirect ophthalmoscopy.

CONCLUSION

This study was concluded that the software method of evaluation of PCO was easy, applicable and more appropriate method to evaluate in clinical studies than any other methods.

REFERENCES

- Gift, B. W., R. V. English, B. Nadelstein, A. K. Weigt and B. C. Gilger, 2009. Comparison of capsular opacification and refractive status after placement of three different intraocular lens implants following phacoemulsification and aspiration of cataracts in dogs. *Vet. Ophthalmol.*, **12**(1): 13-21.
- Raj.M.S., A.R. Vasavada, S.R.K. Johar, V.A. Vasavada and V.A. Vasavada, (2007). Post – Operative Capsular Opacification: A Review, International Journal of Biomedical Science **3**: (237-250).
- Tetz, M.R., G.U. Auffart, M. Sperker, M. Blum and H.E.Volcker, 1997. Photographic image analysis system of posterior capsule opacification. *J.Cataract Refract. Surg.*, **23**(10): 1515-1520.

EFFECT OF SILK FIBROIN LOADED SILVER NANOPARTICLES GEL ON WOUND HEALING IN THREE LABRADOR DOGS

**T.S. Premavathy, Mohamed Shafiuza, R.Siva Shankar,
Mala Shammi, S. Hemalatha**

Department of Veterinary Surgery and Radiology, Madras Veterinary College, Chennai-600007
Tamil Nadu Veterinary and Animal Sciences University

ABSTRACT

Three Labrador retriever dogs (5 months, 1 and 2 years of age) were presented to Madras Veterinary College Teaching Hospital with the history of chronic non-healing open wounds on the left cranial elbow, right dorsolateral abdomen and right ventrolateral abdomen. Physical examination revealed mild yellowish coloured wound with putrid odour and purulent discharge. The Etiology of the wounds was laceration in all the three cases. The subcutaneous tissues were exposed; necrotic tissues and foreign debris were noticed. Silk fibroin loaded silver nanoparticle gel was prepared and applied on days 0, 3, 7, 14, 21 and 28 respectively on the open wound and bandaging done. Silk fibroin is a protein polymer which does not interfere with the host immune system and it can be used in chronic non-healing wounds accelerating wound healing, aids in spreading of keratinocytes and fibroblasts and thereby promoting the growth and development of skin tissue. Clinical examination revealed bright red colour granulation tissue with neovascularisation. On bacteriological examination, *Staphylococcus aureus* and *Pseudococcus aeruginosa* were isolated. Mean per cent epithelialization, wound contraction and wound healing were significantly high.

Keywords: Dog, Chronic non-healing wound, Silk fibroin loaded silver nanoparticle

INTRODUCTION

Wound repair was a complex process requiring coordination of cascade of cellular responses to injury including inflammation, epithelialization, proliferation, angiogenesis and remodelling (Barrientos *et al.* 2008). A major drawback in conventional dressing materials, mainly composed of gauze, is the adherence between the fibers of the gauze materials and the tissue. Separation between the dressing material and the tissue becomes very difficult (Lin *et al.*, 2000). In large wound cases if required donor site is

not available, biomaterials like synthetic or natural polymers and their composites can be used for temporary closure of wounds (Babuet *al.*, 1996). Fibroin protein was known for wound dressing by regulating exudates of wound providing moist environment facilitated re-epithelialization, re-modelling of connective tissues and collagenisation (Teramoto *et al.* 2008). This study is done to assess the clinical outcome of silk fibroin loaded silver nanoparticle gel on wound healing in three Labrador retriever dogs.

Corresponding author: T.S. Premavathy (Pg Scholar),
Email id: premavathytathan@gmail.com, Contact number: 8939172179

MATERIALS AND METHODS

Three Labrador retriever dogs (5 months, 1 and 2 years of age) were presented to Madras Veterinary College Teaching Hospital with the history of chronic non-healing open wounds on the left cranial elbow (Fig1.A), right dorsolateral abdomen (Fig1.B) and right ventrolateral abdomen (Fig1.C). Physical examination revealed mild yellowish coloured wound with putrid odour and purulent discharge. The parameters studied were clinical observation, wound

planimetry, bacteriological evaluation and histopathological analysis on days 0, 3, 7, 14, 21 and 28. Clinical observations were done based on colour, odour and exudate. ABST had taken for the bacteriological evaluation. The silk fibroin loaded silver nanoparticle gel was prepared in the Department of Animal Biotechnology by using silk cocoon and neem leaf extract (Rockwood *et al.* 2011 and Prusty and Parida, 2015). The cases were subjected to the application of Silk fibroin loaded silver nanoparticle gel on subsequent days.



Figure 1. A: Case1- Wound on the left cranial elbow, B: Case2- Wound on right dorsolateral abdomen and C: Case3- Wound on right ventrolateral abdomen.

RESULT AND DISCUSSION

In this study, the colour of the wounds were black with offensive odour and serosanguineous discharge. For bacteriological examination, a sterile cotton swab was rolled over the open wound on days 0, 3, 7, 14, 21 and 28. On days 0 and 3, *Staphylococcus aureus* and *Pseudomonas aerogenosa* were isolated in all the cases. Under sedation, the wounds were debrided and lavaged with sterile normal saline solution. Punch biopsy of 3.5mm sample was taken for histopathological analysis.

Wound planimetry was performed to assess the wound healing on days 0, 3, 7, 14, 21 and 28 by tracing of the wound margins onto a transparent sheet. The percentage of wound epithelialization, wound contraction and total healing were calculated in all the cases. The wounds were applied with silk fibroin loaded silver nanoparticle gel and bandaging was done to immobilise prevent self-mutilation. The bandaging was changed on every 2 days till the complete closure of wound was noticed. The clinical observation of the open wound in case 1 on days 0, 3, 7, 14, 21 and 28 were photographed (Plate 1).

Plate 1: Clinical observation of the open wound – Case 1



Yellow colour appearance of laceration wound - Day 0



Yellow colour with Serosanguineous discharge - Day 3



Presence of Serosanguineous discharge - Day 7



Reduction of wound size - Day 14



Red colour granulation tissue - Day 21



Complete wound healing - Day 28

In case 2 and 3, the clinical observation of open wounds were observed with the foul smelling, necrotic tissues and purulent discharge on day 0. Manual debridement was done under sedation. Other parameters were performed in both the cases and subjected to the application of silk fibroin loaded silver nanoparticles gel on days 0, 3, 7, 14, 21 and 28. Bandaging was done to prevent self-mutilation and movement of the affected region. On days 3, 7 and 14, absence of wound discharge, appearance of granulation tissue and progression of wound contraction around the edges in both the cases. On days 21 and 28, complete epithelialization with granulation tissue formation and complete wound healing were observed in both the cases.

Histopathological Examination

A 3.5 mm punch biopsy instrument was used to take skin specimens from the open wounds on days 0, 3, 7, 14, 21 and 28 with the dogs under general anaesthesia. Punch biopsy used in dogs as a method for skin healing investigation by histological analysis (Hamamoto *et al.*, 2009). The biopsy samples were sent for the histopathological analysis. The specimens were fixed in 10% neutral buffered formalin and processed routinely for histopathological examination. Five micrometer sections were stained with haematoxylin and eosin (H&E) and the progressive decrease in macrophages, fibrosis, and progressive increase in

angiogenesis, epithelialization and collagen level were studied.

The dermis was devoid of epidermal covering and the wound consisted of large defects at the 0th and 3rd day. The wound area was filled with necrotic debris and crust made up of fibrin. The dermis of the wound area in the present study animals showed edema, vascular congestion and showed partial infiltration of neutrophils and macrophages (Plate 2 and 3). On days 7, 14 and 21, the wound area was covered by granulation tissue and fibrin crust, the defect was diminished due to partial contraction of the wound (Plate 4, 5 and 6). There was contraction of fibrin clot, partial re-epithelialization, immature collagenisation and angiogenesis was present.

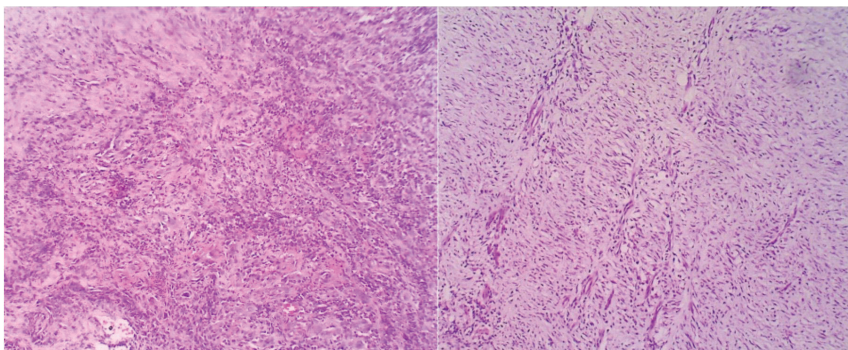
In the present study on 28th day, increased fibrovascular tissue with complete collagenisation, marked angiogenesis and re-epithelialization was observed in all the cases (Plate 7). This treatment promoted healing of chronic wounds by stimulating cellular activities and encompassed good epithelialization with keratinization, matured fibroblast and neovascularisation as opined by Okabayashi *et al.* (2009). Silk protein promoted angiogenesis in all the three cases. The increased vessel growth could be facilitating both the extent and direction of fibroplasia. Improved angiogenesis, therefore, would be contributing significantly to wound healing activity of silk proteins were reported by Padolet *et al.* (2012). Silk fibroin supported adhesion and

growth of anchorage dependent cells such as fibroblasts as opined by Minoura and Aiba (1995). Min *et al.* (2004) reported greater collagen regeneration, less inflammation and less lymphocyte infiltration found in the wound treated with silk film.

Bacterial contamination in a wound significantly impaired healing. The bacterial examination on days 7, 14, 21 and 28 revealed no bacterial organisms in all the cases after the application of silk fibroin loaded silver nanoparticles gel over the wounds. Onget *et al.* (2008) who reported complete destruction of both *Pseudomonas aeruginosa* cultures and *Staphylococcus aureus* cultures with >99.99% efficiency by incorporating silver-enhanced wound dressings *in vitro*.

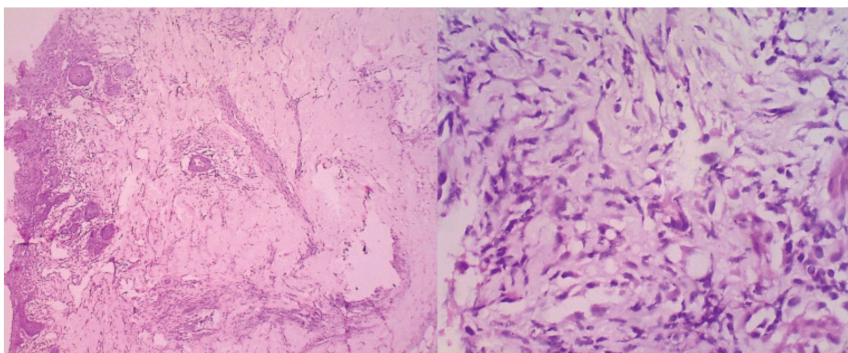
In wound treatment, topical antiseptic application was toxic to fibroblasts and epithelial cells. Conventional dressings, which composed of gauze materials adhered to the tissue and separation of the dressing material became very difficult and interfered with wound granulation and healing process. No such problems were observed in this study for the management of open wounds in the above cases. Hydrogels used in the present study were suitable for cleansing of dry, sloughy or necrotic wounds. They were non-irritant, non-reactive with biological tissue, permeable to metabolites, didn't have residues and able to improve re-epithelialization of wounds. Similar findings were reported by Boateng *et al.* (2008) and Jayakumaret *et al.* (2011).

Histopathological examination of open wounds



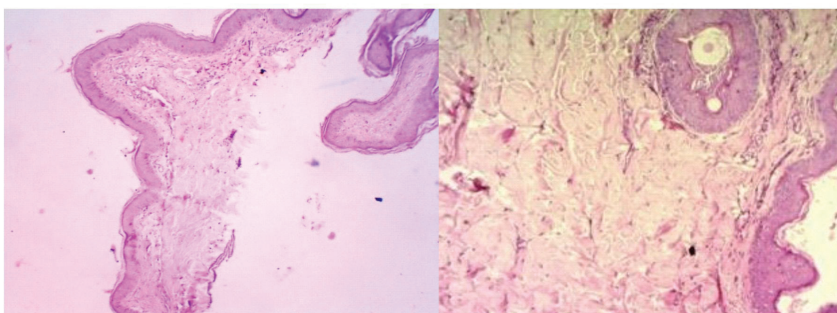
Lack of epithelium, presence of scab and neutrophilic exudate. H&E Stain = 100X (Day 0) - Plate 2

Lack of epithelium, presence of neutrophils with macrophages and mast cells andimmature collagen. H&E Stain = 100X (Day3)-Plate 3



Partial re-epithelialization, mild collagenisation and neutrophilic exudate. H&E Stain = 40X(Day 7) - Plate 4

Partial immature collagenisation and angiogenesis.H&E Stain = 400X (Day 14)- Plate 5



Partial epithelialization and moderate collagenisation.H&E Stain = 100X (Day 21)- Plate 6

Complete re-epithelialization with organised collagenisation. H&E stain = 100 X (Day 28)- Plate 7

CONCLUSION

Silk fibroin loaded silver nanoparticle gel can be a better wound healing biomaterial in dogs. Silk fibroin acted as epidermal cell growth promoter and silver nanoparticles acted as antimicrobial agent leading to faster per cent wound closure. It is non-antigenic, non-toxic, non-irritating with good applicability and adhered well to fibroblasts and epidermal cells. It did not elicit any adverse reaction to the dog's skin in all the three cases. Based on the above observations, it was concluded that, silk fibroin loaded silver nanoparticles treated wounds showed early wound healing and the biomaterial can be safely be applied for cutaneous wound healing in dogs.

ACKNOWLEDGEMENT

The authors acknowledge their gratitude to the Dean and Director of Clinics, Madras Veterinary College, for providing necessary facilities.

REFERENCES

- Barrientos, S., O. Stojadinovic, M. S. Golinko, H. Brem and M. T. Canic, 2008. Growth factors and cytokines in wound healing. *Wound Repair Regen*, **16**: 585-601.
- Babu, P. R., T. P. Sastry, C. Rose and M. N. Rao, 1996. Hydrogels based on gelatin poly (hydroxyethylmethacrylate) and poly (butyl acrylate) graft copolymer impregnated with fibrin. *J. of App. Poly. Sci.* **65**: 555-560.
- Boateng, J.S., K.H. Matthews, H.N. Stevens and G.M. Eccleston, 2008. Wound healing dressings and drug delivery systems: *A review. J. Pharm. Sci.*, **97**: 2892-2923.
- Hamamoto, T., A. Yabuki, O. Yamato, M. Fujiki, K. Misumi, and M. Matsumoto, 2009. Immunohistochemical analysis of cyclooxygenase-2 induction during wound healing in dog skin. *Res. Vet. Sci.* **87**, 349-354.
- Jayakumar, R., M. Prabakaran, P.T. Sudheeshkumar, S.V. Nair, and H. Tamura, 2011. Biomaterials based on chitin and chitosan in wound dressing applications. *Biotechnol. Adv.*, **29**: 322-337.
- Lin, F. H., T. M. Chen, K. S. Chen, T. H. Wu and C. C. Chen, 2000. An animal study of a novel tri-layer wound dressing material-non-woven fabric grafted with N-isopropyl acrylamide and gelatin. *Materials Chem. and Phy.* **64** : 189-195.
- Min, B.M., G. Lee, S.H. Kim, Y.S. Nam, T.S. Lee and W.H. Park, 2004. Electrospinning of silk fibroin nanofibers and its effect on the adhesion and spreading of normal human keratinocytes and fibroblasts *in vitro. Biomaterials.*, **25**:1289-1297.
- Minoura, N. and S. Aiba, 1995. Attachment and growth of cultured fibroblast cells on silk protein matrices. *J. Biomed. Mater. Res.*, **29**(10): 1215-1221.
- Okabayashi, R., M. Nakamura, T. Okabayashi, Y. Tanaka, A. Nagai and K. Yamashita, 2009. Efficacy of

- polarized hydroxyapatite and silk fibroin composite dressing gel on epidermal recovery from full-thickness skin wounds, *J. Biomed. Mater. Res. B. Appl. Biomater.*, **90**(2): 641-646.
- Ong, S., J. Wu, S.M. Moochhala, M. Tan and J. Lu, 2008. Development of a chitosan-based wound dressing with improved hemostatic and antimicrobial properties. *Biomaterials.*, **29**(32): 4323-4332.
- Padol, A.R., K. Jayakumar, N.B. Shridhar, H.D. NarayanaSwamy, K. Mohan and S. Manochaya, 2012. *Int. J. Toxicol. Appl. Pharmacol.*, **2**(3): 31-36.
- Prusty, A. K. and P. Parida, 2015. Development and evaluation of gel incorporated with biogenically synthesised silver nanoparticles. *Journal of Applied Biopharmaceutics and Pharmacokinetics*, **3**: 1-6.
- Rockwood, D. N., C. P. Rucsanda, Y. Tuna, X. Wang, M. L. Lovett and D. L. Kaplan, 2011. Materials fabrication from *Bombyxmorisilk* fibroin. *Nature Protocols*, vol.6 no.10, pp. 1612-1631.
- Teramoto, H., T. Kameda and Y. Tamada, 2008. Preparation of gel film from *B. mori* silk sericin and its characterization as a wound dressing. *Biosci. Biotech. Biochem.* **72**: 3189-3196.

GRANULOSA CELL TUMOUR ALONG WITH PYOMETRA IN A GERMAN SHEPHERD DOG

B. Bibin Becha¹, Sunita Behera¹, M. Navya² and G. Sudha³

¹Ph.D. Scholars, ²Contract Teacher, ³Associate Professor
Department of Veterinary Gynaecology & Obstetrics
Veterinary College, KVAFSU, Hebbal, Bengaluru – 560 024

ABSTRACT

A four year old, nulliparous German Shepherd dog was presented with the history of foul smelling vaginal discharge for the past one month. Abdominal palpation revealed a moderately distended uterus and a hard fluctuating mass in the anterior abdomen. Ultrasonography revealed an oval mass with numerous hypoechoic microcysts and non-uniform echotexture. Anechoic sacculations anterior to the urinary bladder suggestive of pyometra were visible. Ovariohysterectomy was performed as per standard surgical procedures. The left ovary was unusually large, irregular and hard with large number of greyish to brown nodules. Both uterine horns were distended with thickened uterine walls and uneven sacculations. Cut surface of the left ovary showed predominantly cystic parenchyma with fibrin network, filled with serous and blood tinged fluid. Histopathology revealed granulosa cell tumour of the left ovary.

Key words: Granulosa cell tumour, pyometra, dog

INTRODUCTION

Ovarian tumours are uncommon in dogs, accounting for 0.5 to 1.2 per cent of all canine tumours. Granulosa cell tumours (GCT) are sex cord stromal tumours, which accounts for 50 per cent of ovarian tumours and occur in elderly dogs with a median age of 12 years (Klein, 1996). This paper reports a case of unilateral GCT of ovary along with pyometra in a four year old German Shepherd dog.

Case History and Observations

A four year old nulliparous German Shepherd dog was presented to the clinic

with the history of foul smelling vaginal discharge for the past one month. The animal was obese and lethargic. The animal had exhibited oestrus signs three months ago and was not bred in the previous cycles. The owner had presented the animal for neutering.

Abdominal palpation revealed a moderately distended uterus and a hard fluctuating mass in the abdomen. Trans-abdominal ultrasonography revealed a fairly oval structure of 5.4×8 cm in size, having a parenchyma of varying echogenicity with numerous hypoechoic microcysts

Current Postal address: Dr. B. Bibin Becha, Assistant Professor and Head, Livestock Research Station, Thiruvazhamkunnu, P.O., Mannarkkad (Via), Palakkad, Kerala – 678 601, India
e-mail: bibinbecha@gmail.com

(5–15 mm) and non-uniform echotexture. Anechoic sacculations of 2.5 to 3.0 cm diameter anterior to the urinary bladder, suggestive of pyometra was observed. Haematology indicated leukocytosis with thrombocytopenia in the animal.

Based on owner's request for neutering and clinical examination findings, it was decided to perform ovariohysterectomy.

Treatment and Discussion

Ovariohysterectomy was performed as per standard surgical procedures through a midventral abdominal approach under general anaesthesia. The left ovary was unusually large in size with irregular contour (5×8×6 cm) and hard with large number of grey and brown blunt projecting structures. Right ovary was small without any appreciable structures. Both uterine horns were distended (approx. 3 cm in diameter), with thickened uterine walls and uneven sacculations (Fig. 1). Separated left ovary from the bursa was capsulated with small cysts and areas of haemorrhages. Cut surface of the left ovary showed predominantly cystic parenchyma with serous and blood tinged fluids and a few uniform hard structures indicating functional corpora lutea (Fig. 2). Representative tissue samples were collected from left ovary, preserved in 10 per cent formol saline and sent for histopathology.

Histopathology revealed various sized macrofollicular cyst-like structures lined by loosely arranged, multiple layers of granulosa cells. Cysts contained eosinophilic fluid or blood. Masses of granulosa cells were separated by

fibrovascular stroma having lipid containing polygonal theca cells. Neoplastic cells were round or oval, moderately pleomorphic with round hyperchromatic nuclei and scanty eosinophilic cytoplasm. Histopathological features confirmed GCT of left ovary.

GCT originates from ovarian sex cords and may be unilateral or bilateral measuring 4 to 16 cm diameter. The major clinical findings of ovarian tumours are due to the space occupying nature of the intra-abdominal mass, metastatic spread and/or hormonal disturbances. Rarely, GCT may be non-functional and clinical manifestations were absent (Koivisto *et al.*, 2012). Incidence of GCT increases with age, and approximately 20 per cent cases show metastasis. In this case foul smelling vaginal discharge was noticed due to presence of pyometra, which might have produced due to increased production of progesterone from tumour tissues. Occurrence of cystic endometrial hyperplasia or pyometra along with GCT has also been reported (Oliveira *et al.*, 2016).

In the present study, the parenchyma of enlarged ovary was found to be heteroechoic with numerous cysts and had irregular margin, which can be considered as a feature of tumour as observed by Diez-Bru *et al.* (1998) and Kim *et al.* (2012).

Histopathologically, GCT in dogs has various microscopic appearances, such as follicular, cystic or poly cystic, and solid. In this case tumour was macrofollicular, characterized by various sized cystic structures lined with multiple layers of granulosa cells as observed by other researchers (Tavasoli and Solati, 2011).

REFERENCES

- Diez-Bru, N., Garcia-Real, I., Martinez, E.M., Rollan, E., Mayenco, A. and Llorens, P. (1998). Ultrasonographic appearance of ovarian tumors in 10 dogs. *Vet. Radiol. Ultrasound*, 39: 226 – 233.
- Kim, H., Choi, H., Kim, H. and Choi, J. (2012). A giant parovarian cyst in a dog with a granulosa cell tumor. *J. Vet. Med. Sci.*, 74(3): 385 – 389.
- Klein, M.K. (1996). Tumors of the female reproductive system. In: *Small Animal Clinical Oncology*. Eds. S.J. Withrow and E.G. MacEwen. 2nd Edn. W.B. Saunders, Philadelphia. Pp. 347 – 355.
- Koivisto, M.B., Carvalho, I.R., Carreira, T.J., Trevizan, J.T., Machado, G.F., Sueiro, F.A.R., Ciarlini, L.D.R.P. and Gomes, P.B.C.G. (2012). Granulosa cell tumor in a bitch – Case report. In: *Proc. 7th International symposium on Canine and Feline Reproduction*, Whistler, Canada.
- Oliveira, A.R., Flecher, M.C., Jabour, F.F., Souza, T.D., Hardt, I., Vieira, F.T., Rassele, A.C., Vicente, G.C. and Machado, F.M. (2016). Dysgerminoma and granulosa cell tumor in a bitch. *Braz. J. Vet. Pathol.*, 9(1): 31 – 33.
- Tavasoli, A. and Solati, A. (2011). Granulosa cell tumor of the ovary in dog: Case report from Tehran. *J. Cell Anim. Biol.*, 5(4): 66 – 68.

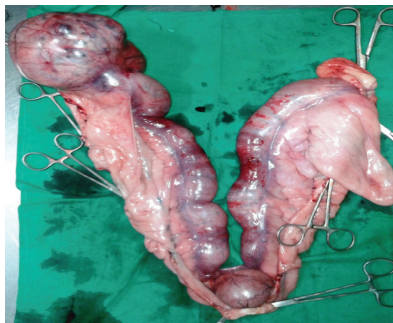


Fig. 1 Gross appearance of pyometra and tumorous left ovary

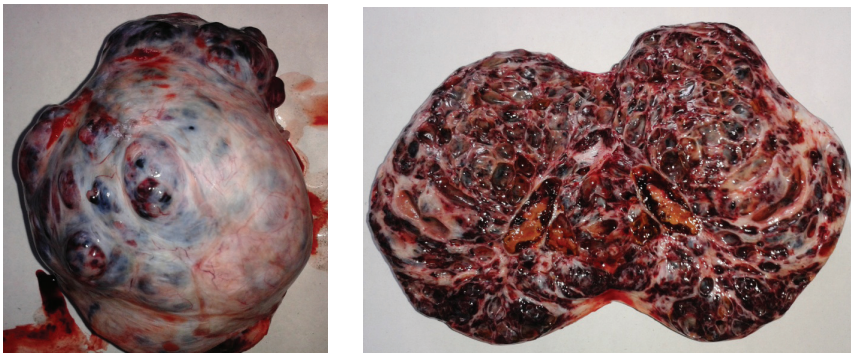


Fig. 2 Gross appearance of tumorous left ovary and its appearance in cut surface

INSTRUCTIONS TO AUTHORS

Scope of the Journal

“**Indian Journal of Veterinary and Animal Sciences Research**” published six times in a year will consider original papers for publication on all aspects of animal and fisheries sciences. The scope of the journal includes animal and fisheries health, management, production and marketing of products. Acceptance of manuscript will be based on scientific merit as judged by referees and Editorial Board.

Submission of manuscripts

Manuscripts should be written in English and the spelling should follow the Oxford English Dictionary. Manuscripts should be submitted in triplicate along with Rs. 500/- as Demand Draft drawn in favour of “**The Editor, IJVASR & Director of Research, TANUVAS, Chennai – 600 051**” as processing fee to the Editor, “**Indian Journal of Veterinary and Animal Sciences Research**”, Directorate of Research and Animal Sciences, Madhavaram Milk Colony, Chennai – 600 051, INDIA. Manuscripts can also be submitted by email to the email id: ijvasr@tanuvas.org.in. Payment can also be made online to the following account.

Account Name: **The Editor, IJVASR & Director of Research, TANUVAS, Chennai**

Account Number: **332902010721641**

IFSC Code: **UBINO533297**

Reg. No.: / Transaction I.D./NEFT :

The authors should give a statement to the effect that the “**Articles sent to IJVASR have not been sent elsewhere for publication**”. The statement should also be signed by all the authors and certified that the work is done as per the mandate of the respective institute. **Email id and contact phone of the first or corresponding author should be provided whereas the authors should submit the copy of the IAEC approval if experimental animals are used.**

Preparation of manuscripts

All manuscripts should be typed on one side of the A4 paper, double-spaced throughout, with margins of at least 25mm all around. All contributions will be subjected to editorial revision.

Major headings are centered, all capitals except for scientific names and the full length papers should consist of Abstract, Introduction, Materials and Methods, Results and Discussion, Acknowledgement (optional) and References. First subheadings begin at the left margin and the text that follows a first subheading should be in a new paragraph.

Full length papers should normally not exceed 3000 words in length including tables and illustrations i.e. approximately five journal pages and should contain the following section, each written concisely:

A **Title** page containing (a) the title of the paper in capital letters in exception for scientific names, (b) the names of authors in full with initials at the beginning, (c) the authors’ department and complete postal address. Superscript numbers should be used to link authors with

other institution. Provide maximum of five key words for full length paper and three for short communication for subject indexing. The author wise contribution should also be mentioned in nutshell.

An **Abstract** will be printed at the beginning of the paper. Abstract should not be more than 150 words emphasizing objectives, experimental procedure, results and conclusions. Use complete sentences and limit the use of abbreviations. It should be in a form suitable for abstracting journals to use.

A brief **introduction** with specific emphasis on the necessity for such a kind of research may be given.

Materials and methods section may refer to previous description of methods whenever possible. This section should include experimental designs and methods of statistical analysis.

Results and Discussion may contain subheading if appropriate. This part should be brief and to the point, without repetition of results.

An **Acknowledgement** section, if required, may be given.

References section should contain only essential references which should be listed alphabetically and written as indicated below. In the text, give the author's name followed by the year in parenthesis: Suresh (2009). If there are two authors, use 'and': Suresh and Mani (2015); but if cited within parenthesis: (Suresh and Mani, 2015). When reference is made to a work by three or more authors, the first name followed by et.al. should be used: Rama et.al.(2015); but if cited within parenthesis: (Rama et.al., 2015). Reference to unpublished data and personal communications should not appear in the list but should be cited in the text only (e.g. Amutha T, 2015. Unpublished data).

Journal articles and abstracts

Bardbury, J.M., Mc Carthy, J.D and Metwali, A.Z. (1990). Micro immunofluorescence for the serological diagnosis of avian Mycoplasma infection. *Avian Pathology*, **19**:213-222.

Raja, S., Rani, A., Ravi, M and Kumar. K. (2007). Histopathology of CPV infection. Page no. 120-122....Venue...Date...Place...

Books and articles within edited books

Rundall, C.J. (1991). A colour Atlas of Diseases of the Domestic Fowl and Turkey. 2nd ed. London. Wolf Publishing Ltd. 175 p.

Handbooks, Technical bulletins, Thesis and Dissertations

Callow, L.L and Dalgliesh, R.J. (1982). Immunity and Immunopathology in Babesiosis. In: S. Choen and K.S. Warren (Ed) Immunology of Parasitic Infections. Blackwell, Oxford. pp 475-526.

Electronic publications

Tables should be typed on separate sheets, numbered consecutively in Arabic Numerals and have a short descriptive heading. Units of measure for each variable measured should be indicated. Appropriate estimates of variation (Standard error, standard deviation) must be provided with means. Use superscript letters for the separation of means in the body of the table and explain these in footnotes.

Illustrations, referred to as “figures” (Fig. 1 etc.) should be on separate sheets and submitted larger than the size desired for reproduction. Information in tables must not be duplicated in figures and vice versa. Legends, should be provided for each illustration. Line drawings should be either in black ink on smooth white paper or thin board or a good quality laser printout. Photographs and photomicrographs should be printed on glossy paper with good contrast. Magnification for photomicrographs should be indicated. All illustrations should bear on the reverse side, the first author’s name and the figure number, the ‘top’ of the figure should be indicated. While sending the manuscripts in email, and the figures should be separately sent in JPEG format but for gel pictures it should be in TIFF format with good resolution.

Short communications and Case Reports should have a title page as described for full length papers and should comprise approximately 1000 words including tables, illustrations and references. They may contain not more than two tables or illustrations. Methods, results and discussion should be in single section without headings. References should be kept to a minimum and be in the form described above.

Review should have a title page as described for full length papers and should contain approximately 4000 words including tables, illustrations and references.

Units, symbols and abbreviations

Units should conform to the International System of Units (refer Baron, D.N. (1994). Units, Symbols and Abbreviations: A Guide for Biological and Medical Authors. 4th ed. London. Royal Society of Medicine). Abbreviations should not be used in the title, section heading or at the beginning of sentences. As a rule, author-coined abbreviations should be in all capital letters. These should be spelled out in full with the abbreviation following in parentheses the first time they are mentioned.

Proofs

Proofs will usually be sent to the first or corresponding author. Only typesetter’s errors may be corrected; no changes in, or additions to, the edited manuscript will be allowed. It is a condition of acceptance that the Editors reserve the right to proceed to press without submitting the proofs to the author. While reasonable care will be taken to ensure that proof reading is correctly done, neither the Editors nor the Publishers shall be responsible for any errors.

Reprints

It has been decided to discontinue the supply of 25 reprints as the contents of the articles is hosted as PDF in TANUVAS website. (www.tanuv.ac.in/ijvasr.html).

Rejected article

Hard copy of the rejected articles will not be referred to the authors. The chief editor has the sole rights to either accept or reject the manuscripts based on their merits without reasoning.

FORM IV (See Rule 8)

1. Place of Publication : University Publication Division (Printing Press)
Tamil Nadu Veterinary and
Animal Sciences University,
Madhavaram Milk Colony, Mathur Road,
Chennai – 51. Ambattur Taluk
Thiruvallur District
2. Periodicity of Publication : Bi-Monthly
3. Printer's Name : **Dr. S.A. Asokan**
Whether citizen of India Yes
Address Director of Distance Education
Tamil Nadu Veterinary and
Animal Sciences University,
Madhavaram Milk Colony
Chennai – 51. Ambattur Taluk
Thiruvallur District
4. Publisher's Name : **Dr. T.J. Harikrishnan**
Whether citizen of India Yes
Address Director of Research
Tamil Nadu Veterinary and
Animal Sciences University,
Madhavaram Milk Colony
Chennai – 51. Ambattur Taluk
Thiruvallur District
5. Chief Editor's Name : **Dr. S. Thilagar**
Whether citizen of India Yes
Vice-Chancellor
Tamil Nadu Veterinary and
Animal Sciences University,
Madhavaram Milk Colony, Chennai – 600 051.
6. Name and address of individuals : **The Registrar**
who own the newspaper and parents
or share holders holding more than
one per cent of the total capital Tamil Nadu Veterinary and
Animal Sciences University,
Madhavaram Milk Colony, Chennai – 600 051.

I, Dr. T.J. Harikrishnan hereby declare that the particulars given are true to the best of my knowledge and belief.

Dr. T.J. Harikrishnan
Signature of Publisher

All the contributing authors are requested to bestow their personal attention while submitting the revised manuscripts for spelling mistakes and correctness of language.

Chief Editor

The Indian Journal of Veterinary and Animal Sciences Research is indexed in the abstracting journals of the CAB International, Zoological Abstracts of Web of Knowledge published by Thomson Reuters and Indian Science Abstracts published by NISCAIR, India.